

ORIGINAL INVESTIGATION

Open Access



Circulating Receptor Activator of Nuclear Factor kB Ligand and triglycerides are associated with progression of lower limb arterial calcification in type 2 diabetes: a prospective, observational cohort study

Olivier Bourron^{1,2,6,18,21*} , Franck Phan^{1,2,6,18}, Mamadou Hassimiou Diallo³, David Hajage¹⁹, Carole-Elodie Aubert², Aurélie Carlier^{1,2,6}, Joe-Elie Salem^{1,4,5,6}, Christian Funck-Brentano^{1,4,5,6}, Salim Kemel^{1,7,8}, Philippe Cluzel^{1,6,7,8}, Alban Redheuil^{1,6,7,8}, Jean-Michel Davaine¹, Ziad Massy⁹, Romuald Mentaverri¹⁰, Dominique Bonnefont-Rousselot^{11,12}, Philippe Gillery^{13,14}, Stéphane Jaisson^{13,14}, Cees Vermeer¹⁵, Jean-Marc Lacorte^{1,16,17}, Nesrine Bouziri²⁰, Suzanne Laroche^{1,2,6}, Chloé Amouyal^{1,2,6} and Agnes Hartemann^{1,2,6,18}

Abstract

Background: Lower limb arterial calcification is a frequent, underestimated but serious complication of diabetes. The DIACART study is a prospective cohort study designed to evaluate the determinants of the progression of lower limb arterial calcification in 198 patients with type 2 diabetes.

Methods: Lower limb arterial calcification scores were determined by computed tomography at baseline and after a mean follow up of 31.20 ± 3.86 months. Serum RANKL (Receptor Activator of Nuclear factor kB Ligand) and bone remodeling, inflammatory and metabolic parameters were measured at baseline. The predictive effect of these markers on calcification progression was analyzed by a multivariate linear regression model.

Results: At baseline, mean \pm SD and median lower limb arterial calcification scores were, 2364 ± 5613 and 527 respectively and at the end of the study, 3739 ± 6886 and 1355 respectively. Using multivariate analysis, the progression of lower limb arterial log calcification score was found to be associated with (β coefficient [slope], 95% CI, p-value) baseline log(calcification score) (1.02, 1.00–1.04, $p < 0.001$), triglycerides (0.11, 0.03–0.20, $p = 0.007$), log(RANKL) (0.07, 0.02–0.11, $p = 0.016$), previous ischemic cardiomyopathy (0.36, 0.15–0.57, $p = 0.001$), statin use (0.39, 0.06–0.72, $p = 0.023$) and duration of follow up (0.04, 0.01–0.06, $p = 0.004$).

Conclusion: In patients with type 2 diabetes, lower limb arterial calcification is frequent and can progress rapidly. Circulating RANKL and triglycerides are independently associated with this progression. These results open new therapeutic perspectives in peripheral diabetic calcifying arteriopathy.

Trial registration NCT02431234

*Correspondence: olivier.bourron@aphp.fr

²¹ Diabetology Department, Pitié-Salpêtrière Hospital, 47-83 Boulevard de l'Hôpital, Paris, France

Full list of author information is available at the end of the article



Keywords: Arterial calcification, Biomarkers, Calcium score, Mediacalcosis, Peripheral artery disease, Type 2 diabetes

Background

Arterial calcification is particularly prevalent in ageing, diabetes or renal failure [1]. Lower limb arterial calcification is observed in up to 50% patients with type 2 diabetes and is independently associated with cardiovascular morbidity, renal failure and mortality [2–4]. Due to its implication in peripheral arterial occlusive disease (PAOD), a major complication of lower limb arterial calcification is amputation [4–6]. Although endovascular and open surgery are efficient procedures in diabetic PAOD, arterial calcification is a risk factor of arterial revascularization failure [7, 8]. Understanding the pathophysiology of the progression of peripheral arterial calcification is important in opening new therapeutic perspectives.

For a long time, the mechanism of arterial calcification was considered as a passive process resulting in the precipitation and deposition of hydroxyapatite crystals. But now vascular calcification is considered as a complex and actively controlled molecular process of “arterial ossification”, involving the differentiation of vascular smooth muscle cells and other cells into osteoblast-like cells [4, 9, 10]. There is growing evidence that the OPG (osteoprotegerin)/RANKL (Receptor Activator of Nuclear factor κ B ligand) signaling axis could also play a central role [11, 12]. Others pathways are also suspected such as i) mechanical stress and hypertension, ii) induction of calcification by lipids, inflammation, glucose, advanced glycation end-products (AGEs) and their soluble receptor (sRAGE) [9, 13–15]. Furthermore, neuropathy could also participate in the calcification process [4].

The prospective DIACART study was designed to look at the potential determinants of lower limb arterial calcification progression in type 2 diabetes. The primary objective was to determine if the RANKL pathway is associated with the progression of lower limb arterial calcification in Type 2 Diabetes. Secondary objectives were to assess association between the others proteins of bone remodeling, traditional cardiovascular risk factors, markers of inflammation, metabolic parameters and the progression of lower limb arterial calcification.

Methods

Study design

The DIACART (for “DIAbète et Calcification ARTerielle”) study is a prospective monocentric cohort study [4]. The recruitment period extended from February to October 2014. Inclusion criteria were: Type 2 Diabetes with at least a history of coronary artery disease and/or peripheral arterial occlusive disease and/or age > 50 years

for men and > 60 years for women. Exclusion criteria were: [1] an estimated glomerular filtration rate (GFR) calculated by the modification of diet in renal disease equation < 30 ml/min, [2] a history of lower limbs angioplasty and/or bypass, or [3] type 1 diabetes. The study was approved by the local ethics committee and registered in ClinicalTrials.gov (Identifier: NCT02431234). All patients were informed of the study objectives and procedures. Participants gave their written informed consent of participation prior to inclusion.

Study follow-up

At the inclusion visit and after a mean follow up of 31.2 ± 3.9 months (median 30.5 months and range 26–44.8 months), all patients had a clinical evaluation, laboratory blood tests, and helicoidal CT (computerized tomography) scans. Patient interviews focused on comorbidities and their personal disease history. Their medical records were reviewed to check the clinical information and to record concomitant treatments. Ischemic cardiomyopathy was defined as a history of myocardial infarction, acute coronary syndrome or any surgical procedures undergone due to coronary artery disease.

Assessment of peripheral neuropathy

The physician who performed the clinical tests was not informed of the calcification score nor the laboratory results. Peripheral neuropathy was assessed by the Neuropathy Disability Score (NDS) [16]. It assesses sensory vibration of the big toe using a 128-Hz tuning fork, temperature sense on the dorsum of the foot using a tuning fork with a beaker of ice or warm water, pinprick sense by applying a pin near the nail of the big toe and the Achilles reflex test. Each sensory test scores 0 for normal and 1 for abnormal sensation, on each foot. Achilles reflexes score 0 if they are present, 1 if they are present with reinforcement and 2 if they are absent, for each foot. An $NDS \geq 6$ leads to the diagnosis of diabetic peripheral neuropathy.

Laboratory evaluations

Blood and urine samples were collected after an overnight fast for the measurement of routine biochemistry diagnostic tests, glucose, glycated hemoglobin (HbA1c), cholesterol (total and HDL), triglycerides, high-sensitivity C-reactive protein (hsCRP), calcium, phosphate, urine albumin and creatinin.

Total OPG was measured by ELISA (ELISA MicroVue; Quidel Corporation), and soluble RANKL (sRANKL) concentrations were evaluated using the human RANKL

Single Plex kit from Millipore (Ref: HBN51K1RANKL; eBioscience). With this kit, the minimum detectable concentration of sRANKL is 4.8 pg/ml and inter- and intra-assay precision is below 6% coefficient of variation. The OPG to sRANKL concentration ratio was calculated for each patient without any postanalytical modifications.

Circulating total adiponectin concentrations were measured in serum using an enzyme-linked immunosorbent assay kit (ALPCO, Eurobio, Paris, France) as recommended by the manufacturer, with the lowest detection limit of 0.4 µg/ml for total adiponectin. Inter-assay coefficients of variation of low and high human pool controls for total adiponectin were 7.93% and 8.46%, respectively.

Selected assays (including desphospho-uncarboxylated MGP (dp-ucMGP) and total-uncarboxylated MGP (t-ucMGP) assays) were performed on thawed samples, which had been frozen and stored at -80 °C. A dual-antibody ELISA was used to measure dp-ucMGP levels; the capture antibody was directed against the non-phosphorylated MGP sequence 3–15 (mAb-dpMGP; VitaK BV, Maastricht, Netherlands) and the detecting antibody was directed against the uncarboxylated MGP sequence 35–49 (mAb-ucMGP; VitaK BV). The same antibodies have already been used elsewhere for immunohistochemical staining [15, 17]. Intra-assay variability was 5,6% for dp-ucMGP and 8,9% for t-ucMGP, while inter-assay variability was 9,9% for dp-ucMGP and 11,4% for t-ucMGP. A competitive (single-antibody) ELISA was used to measure t-ucMGP levels, as described previously [18, 19].

Soluble RAGE (sRAGE) was measured on plasma samples with a commercially available ELISA kit (Quantikine Human RAGE Immunoassay, R&D Systems, Minneapolis, USA; see <http://www.rndsystems.com/Products/DRG00> for detailed description of the measurement method).

Advanced Glycation End products (Carboxymethyllysine (CML), Methylglyoxal-derived hydroimidazolone1 (MG-H1) and pentosidine) were determined on serum samples by liquid chromatography coupled with tandem mass spectrometry (API4000 system ABSciex, Les Ulis, France) [20]. AGEs concentrations were expressed as a ratio of total protein concentrations.

Total cholesterol and triglyceride concentrations were determined by an automated enzymatic method (Konelab, Thermoclinical LabSystems, Cergy Pontoise, France and Biomerieux, Marcy L'Etoile, France, respectively). HDL-cholesterol was measured by a direct method (Konelab). LDL-cholesterol was calculated using Friedewald's equation on triglycerides ≤ 3.9 mmol/l or directly measured when triglycerides were > 3.9 mmol/l using a Konelab kit. Inter- and intra-assay coefficients of variation were 2.2% and 0.9%, 1.3% and 0.9%, 3.5% and 0.97%, 1.3%

and 0.9%, respectively for total cholesterol, HDL-cholesterol, triglycerides and LDL-cholesterol, respectively.

Serum human Fetuin A was evaluated using ELISA kit (TECOmedical Group, France) and was performed according to instructions provided by Epitope Diagnostics (intra-assay coefficient of variation: $< 5.5\%$, inter-assay coefficient of variation: $< 6.8\%$; detection limit of the assay: 5.0 ng/ml).

Serum human C-terminal FGF23 (Fibroblast Growth Factor 23) was determined using an ELISA kit (TECOmedical Group, France) and was performed according to the instructions provided by Immotopics (intra-assay coefficient of variation: $< 2.4\%$, inter-assay coefficient of variation: $< 4.7\%$; detection limit of the assay: 1.5 RU/ml).

IL-6 (Interleukine-6) was quantified using automated assays with Access 2 (Beckman Coulter Inc, Villepinte, France) according to the manufacturer's instructions. IGF-1 (Insulin like Growth Factor 1), 25-hydroxyvitamin D, intact parathyroid hormone (iPTH) were measured by a single step chemiluminescence sandwich method on the Liaison XL (DiaSorin) analyzer. Because IGF-1 decreases with age, a standardized IGF-1 score was calculated as previously described [IGF-1 score = $(\log [\text{IGF-1 (micrograms per liter)}] + 0.00625 \times \text{age} - 2.555) / 0.104$] [21].

Assessment of insulin resistance

Insulin resistance was evaluated by the TyG index (triglycerides glucose index), which was calculated as $\ln(\text{fasting triglycerides [mg/dL]} * \text{fasting glucose [mg/dL]} / 2)$ [22].

Imaging for below-knee arterial calcification score

Below-knee artery calcification score was obtained after scanning with a 128-slice multidetector dual source CT-scanner (Somatom Definition Flash, Siemens Healthineers Healthcare, Erlangen, Germany) without contrast, from the base of the patella down to the ankle. Three millimeter cross-sectional slices were analyzed. The analysis was performed by radiologists who were blinded to the results of color duplex ultrasonography, laboratory tests or clinical examination, using a commercially available software package (Heartbeat CaScore, Philips Healthcare, Eindhoven, Netherlands). On cross-sectional images, areas of calcification along below-knee arteries with a density of ≥ 130 Hounsfield units attenuation and a surface area of $> 1 \text{ mm}^2$ were identified semi automatically. Calcification score, determined according to the method described by Agatston et al., was obtained separately for each of the main below-knee arteries (distal popliteal, anterior tibial, posterior tibial and peroneal arteries) and then added up to obtain the total calcification score [23]. Below-knee artery calcification scoring was performed at the inclusion visit and at the end of the study [4].

Statistical analysis

Quantitative variables were described by their mean, standard deviation, median and quartiles Q1–Q3. Qualitative variables were described by their frequency and percentage. The effect of baseline serum RANKL on the progression of calcification of leg arteries between baseline and the end of the follow-up was analyzed using univariable and multivariable linear regression model adjusted on arterial calcification score at baseline. This method is fully equivalent to an analysis of covariance (ANCOVA). Distributions of biological parameters were checked graphically and those with a log-normal distribution were subsequently transformed before any analysis to improve normality. This model was adjusted on baseline cardiovascular risk factors and other factors known to be associated with vascular calcification (age, gender, tobacco use, hypertension, waist circumference, BMI, triglycerides, total cholesterol, HDL-cholesterol, LDL-cholesterol, ApoA1, urinary albumin-creatinine ratio, HbA1c, lower limb log calcification score at baseline, hsCRP, parathormone, glomerular filtration rate (MDRD), and duration of follow up). Cook distance was used for the detection of highly influential observations on the coefficient estimates. A backward stepwise variable selection procedure based on the Akaike Information Criterion was used to select the final multivariate model. Coefficient estimates were provided with their corresponding 95% confidence intervals.

The effects of other bone remodeling factors, markers of inflammation and glycation (Calcium, iPTH, OPG, MGP, IGF-1, Fetuin A, FGF-23, hsCRP, IL-6, carboxymethyllysine, MG-H1, pentosidine, RAGE) were evaluated with multivariate linear regression model using the same procedure.

Significance was defined as p-values of less than 0.05. Statistical analyses were performed using R 3.5.1 (<http://www.R-project.org>).

Sample size

It was assumed that the correlation between the logarithm of the serum RANKL level and the logarithm of the artery calcification score at 2 years, adjusted on the covariates, is about 0.20 (or an R^2 of 0.04). In addition, it was assumed that the multivariate linear model (based on RANKL and associated risk factors) will explain about 15% of the variability. Under these assumptions, at least 169 patients were needed to demonstrate a significant effect of RANKL on the calcification score with an alpha risk of 5% (bilateral formulation) and a power of 80%. In order to take into account 15% of patients lost to follow-up, 198 subjects were planned for the study.

Results

At baseline, a total of 198 patients were included in the DIACART study. During the study follow-up (mean 31.2 ± 3.9 months; median 30.5 months and range 26–44.8), 18 patients were lost to follow up and 11 patients died before the second determination of calcification score. RANKL measurements were missing in 6 patients. This led to 163 evaluable patients.

Patients' characteristics

The characteristics of the participating patients at baseline ($n=163$) are presented in Table 1. At baseline, patients were predominantly middle aged (median [Q1–Q3], 65 years [58–70]) overweight (median [Q1–Q3] BMI, 28.39 kg/m² [25.44;31.77]) male (80%), with a long duration of diabetes (median [Q1–Q3], 12 years [6–20]), and a relatively good glycemic control (median [Q1–Q3] HbA1c, 7.4% [6.9;8.25]; 57 mmol/mol [52;67]). Less than 20% of the patients had laserized retinopathy or neuropathy. Nearly a third of the patients had nephropathy but the GFR (median [Q1–Q3], 77 ml/min [63.5;89]) was relatively preserved as expected since a GFR of <30 ml/min was an exclusion criterion. Most of the patients had ischemic cardiomyopathy (69%), were smokers (60%) and treated for hypertension (80%) with arterial pressure at target (126 ± 17 mmHg). They often took statins (90%), antiplatelet therapies (83%), β -blockers (63%) and renin-angiotensin system inhibitors (79%). Diabetes was often treated with metformin, insulin and sulfonylurea (respectively 79%, 57% and 46%).

Progression of lower limb calcification and independent predictors

At baseline, mean \pm SD and median lower limb arterial calcification scores were 2364 ± 5613 and 527 respectively. At the end of the study, mean \pm SD and median arterial lower limb calcification scores were 3739 ± 6886 and 1355 respectively. In Table 2, lower limb arterial calcification scores are shown for all patients and according to circulating RANKL quartiles at baseline.

In univariate analysis (cf Table 3), lower limb log calcification score at the end of follow-up is correlated with (β coefficient [slope], 95% CI, p-value) baseline log calcification score (0.88, 0.81–0.95, $p < 0.001$), age (0.15, 0.06–0.25, $p < 0.001$), male sex (3.51, 1.64–5.38, $p < 0.001$), tobacco use (for patients without tobacco consumption, -1.69 ; -3.27 to -0.12 , $p = 0.0035$), ischemic cardiomyopathy (3.73, 2.15–5.32, $p < 0.001$), MDRD (-0.06 , -0.10 to -0.02 , $p = 0.005$), t-ucMGP (-2.74 , -4.85 to -0.64 , $p = 0.011$), dp-ucMGP (1.61, 0.53–2.70, $p = 0.004$), Fetuin A (-1.82 , -3.36 to -0.29 , $p = 0.02$), pentosidine (2.16, 0.35–3.96, $p = 0.02$), CML (1.61, 0.53–2.70, $p = 0.004$),

Table 1 Baseline characteristics of the patients (n = 163)

Age (years)	65 [58–70]
Sex male n (%)	130 (80)
BMI (kg/m ²)	28.39 [25.44;31.77]
Waist circumference (cm)	101 [93;110]
Smoking habit (active or past) n (%)	97(60)
Diabetes duration (years)	12 [6–20]
Glycemia (mmol/l)	7.7 [6.2;9.25]
HbA1c % (mmol/mol)	7.4 [6.9;8.25] (57 [52;67])
Ischemic cardiomyopathy n (%)	117 (69)
Calcification score	526.73 [55.25–2253.41]
NDS ≥ 6 n (%)	25 (15)
Laserised retinopathy n (%)	26 (16)
eGFR MDRD (ml/min)	77 [63.5;89]
Microalbuminuria (mg/l)	20 [8.55;66.95]
urinary albumin-creatinine ratio > 3 mg/mmol n (%)	53 (33)
Hypertension n (%)	131 (80)
Triglycerides (mmol/l)	1.25 [0.91;2]
Total Cholesterol (mmol/l)	3.52 [2.7;4.31]
HDL Cholesterol (mmol/l)	1.06 [0.88;1.27]
LDL Cholesterol (mmol/l)	1.84 [1.46;2.35]
SBP (mmHg)	124 [115;135]
DBP (mmHg)	72 [67;78]
hsCRP (mg/l)	1.20 [0.5;2.6]
IL-6 (pg/ml)	2.80 [2.1;4]
Adiponectine	3.50 [2.7;5.05]
TyG index	8.99 [8.53;9.49]
IGF-1 (ng/ml)	139 [105;170]
Standardized IGF-1 (ng/ml)	−0.12 (−1.21 to 0.8)
MG-H1 (μmol/g prot)	2.79 [2.48;3.06]
Pentosidine (nmol/g prot)	1.10 [0.84;1.42]
Carboxyméthyllysine (μmol/g prot)	0.14 [0.13;0.16]
sRAGE (pg/ml)	828.77 [582.61;1181.96]
Corrected calcium (mmol/l)	2.31 [2.24;2.38]
Phosphorus (mmol/l)	1.01 [0.91;1.13]
iPTH (pg/ml)	47.05 [36.75;64.97]
sRankl (pmol/l)	8.06 [2.59;16.8]
OPG (pmol/l)	6.06 [5.26;7.13]
OPG/RANKL	0.73 [0.33–2.25]
Fetuin A (g/l)	0.65 [0.51;1.13]
FGF-23 (U/ml)	20.87 [11.65;34.07]
t-ucMGP	4733 [3680–5490]
Dp-ucMGP	561 [339–761]
Insulin use n (%)	75 (46)
Metformin use n (%)	130 (80)
Sulfonylurea use n (%)	71 (43)
Statin use n (%)	146 (90)
Antiplatelet use n (%)	136 (83)
ARB and ACE inhibitors use n (%)	128 (79)
Beta-blocker use n (%)	102 (63)

Table 1 (continued)

Data are given as median (and quartiles Q1–Q3) or as the number (and percentage) for binary variables

ARB angiotensin receptor blockers, ACE angiotensin converting enzyme, BMI body mass index, CVD cardiovascular disease, DBP diastolic blood pressure, FGF-23 fibroblast growth factor 23, eGFR MDRD estimated glomerular filtration rate calculated with the modification of diet in renal disease formula, HbA1c haemoglobin A1C, HDL high density lipoprotein, hsCRP high sensitivity C-reactive protein, IGF-1 insulin growth factor-1, IL-6 interleukin 6, iPTH intact parathyroid hormone, LDL low density lipoprotein, MG-H1 Methylglyoxal-derived hydroimidazolone1, dp-ucMGP desphospho-uncarboxylated matrix gla protein, t-ucMGP Total uncarboxylated matrix gla protein, NDS neuropathy disability score, OPG osteoprotegerin, SBP systolic blood pressure, sRAGE soluble form of receptor for advanced glycation end products, sRANKL soluble form of Receptor Activator of Nuclear factor κB Ligand, TyG triglyceride glucose index

MG-H1 (−3.15, −5.79 to −0.51, $p=0.020$) and iPTH (0.04, 0.01–0.07, $p=0.012$). But peripheral neuropathy (defined by $NDS \geq 6$) was not associated with lower limb arterial calcification progression (0.15, −0.86–1.17, $p=0.765$). HbA1c, diabetes duration, calcemia, phosphoremia, RAGE, Fetuin A, FGF-23, IGF-1, LDL-cholesterol, HDL-cholesterol, total cholesterol, blood pressure, BMI, hsCRP and IL-6 were also not associated with lower limb arterial calcification progression.

In multivariate analysis (cf Table 4), and after exclusion of highly influential observations, progression of lower limb arterial calcification was associated with (β coefficient [slope], 95% CI, p -value) baseline log calcification score (1.02, 1.00–1.04, $p<0.001$), duration of follow up (0.04, 0.01–0.06, $p=0.004$), ischemic cardiomyopathy (0.36, 0.15–0.57, $p=0.001$), statin use (0.39, 0.06–0.72, $p=0.023$), triglycerides (0.11, 0.03–0.20, $p=0.007$) and log RANKL (0.07, 0.02–0.11, $p=0.016$). Similarly, the analysis of the predictive power of the others biomarkers showed that Log(OPG/RANKL ratio) was also associated with lower limb arterial calcification progression (−0.06, −0.12–0, $p=0.03$) while OPG alone (−0.15, −0.52–0.23, $p=0.409$) was not. Other markers potentially involved in arterial calcification pathophysiology were not implicated in lower limb arterial calcification progression in the DIACART study.

Discussion

Our study confirms that the prevalence of arterial calcification in the lower limbs of patients with type 2 diabetes and with a high cardio-vascular risk is high, but also shows that this pathological process progresses significantly. The association of the calcification progression with the severity of pre-existing calcification, the duration of follow-up and a history of ischemic cardiomyopathy has already been described. But our results highlight the putative role of the RANK/RANKL pathway and circulating triglycerides in this phenomenon.

Table 2 Calcium score at baseline and at the end of the follow-up in all the population and according to circulating RANKL quartiles (n = 163)

	All	sRANKL quartiles			
		< Q1	[Q1–Q2]	[Q2–Q3]	> Q3
Calcium score at baseline	527 [55–2253]	1200 [152–2311]	535 [66–1339]	384 [237–3175]	815 [55–4488]
Calcium score at the end of follow-up	1355 [167–4235]	2204 [410–4084]	1355 [133–3393]	1234 [404–5701]	1840 [153–7265]

Data are given as median (and quartiles Q1–Q3). sRANKL soluble form of Receptor Activator of Nuclear factor κB Ligand

RANKL/RANK/OPG system and lower limb arterial calcification

Our results suggest that an emerging pathway in arterial calcification pathophysiology is the RANKL/RANK/OPG system. RANKL, which is produced by osteoblasts, vascular cells, stromal cells, T-cells, macrophages and monocytes, binds to its receptor RANK which is expressed by osteoclasts but also by vascular smooth muscular cells (VSMCs). This binding activates several downstream targets such as the Nuclear Factor-κB (NFκB). In contrast, OPG acts as a decoy receptor blocking the RANKL-RANK interaction [24]. RANKL actively promotes vascular calcification by inducing, via its receptor RANK, differentiation of VSMCs into osteoblast-like cells [12, 25]. RANK is a member of the Tumor Necrosis Factor Receptor (TNFR) superfamily, and TNFR-1 is associated with the risk of major amputation in diabetes [26]. OPG was shown to be associated with carotid intima-media thickness in T2D patients [27] and is also suspected to be involved in the regulation of the vascular calcification process. Indeed, OPG-deficient mice develop accelerated arterial calcification whereas inactivation of RANKL signaling in these mice counteracts this effect [28]. This suggests that RANKL plays a central role in the regulation of vascular calcification, whereas OPG would rather play a modulatory function. In human, both tissue and serum RANKL have been associated with lower limb artery and carotid calcification [12, 29]. The RANKL/OPG ratio has also been correlated with the coronary artery calcium score [30]. In the DIACART study, both circulating RANKL and RANKL/OPG ratio, but not serum OPG, were correlated with the progression of lower limb arterial calcification. Thus when circulating RANKL concentration increases, it would be associated with an acceleration of the arterial calcification process.

Lipids and lower limb arterial calcification

We also found an association between plasma triglycerides and the progression of lower limb arterial calcification. However we did not find any correlation of the calcification progression with other lipids, although sortilin, which is implicated in lipid metabolism,

extra-cellular matrix mineralization and atherosclerotic plaque burden, was recently found to be associated with the presence of PAOD in T2D [31]. Previous studies have shown a relationship between triglycerides and coronary artery calcium score progression in patients with or without diabetes [32, 33]. When accumulated in the tissue, triglycerides can generate species such as free fatty acids, sphingolipids and particularly ceramides, which could induce pathophysiological pathways involved in vascular calcification. Saturated fatty acids are associated with vascular disease and with the development of vascular smooth muscle cell calcification via the NF-κB pathway induction [34]. Among sphingolipids species, ceramides are known to induce human vascular smooth muscle cell calcification via p38 mitogen-activated protein kinase signaling [35]. It would be interesting to study the role of lipid derivatives, such as sphingolipids, on lower limb vascular calcification development occurring in type 2 diabetes.

Insulin resistance and lower limb arterial calcification

Circulating triglycerides are also a strong marker of insulin resistance and visceral obesity. However in our study other biomarkers of insulin resistance and/or visceral obesity (Triglyceride-Glucose index (TyG index), waist circumference and serum adiponectin) did not correlate with the progression of lower limb vascular calcification. This suggests that triglycerides act through an independent pathway to insulin resistance and visceral obesity.

Inflammation and lower limb arterial calcification

Recently low levels of Omentin-1, which is an adipokine with anti-inflammatory properties, were shown to be associated with the presence and severity of PAOD in T2D [36]. However inflammatory markers like IL-6 and hsCRP were not associated with the progression of the calcification in our study.

Therapeutic perspectives

Denosumab is a human monoclonal antibody which inhibits the RANKL pathway. It is one of the latest therapeutic options for osteoporosis [37]. It has been

Table 3 Univariate linear regression analysis: variables associated with calcification score at the end of follow-up (n = 163 patients; mean follow-up duration = 31.20 ± 3.86 months)

	β coefficient [slope]	IC 95	p value
Log(calcium score at baseline)	0.88	[0.81;0.95]	< 0.001
Follow-up duration	-0.07	[-0.27;0.13]	0.49
Age	0.15	[0.06;0.25]	0.001
Sex (male)	3.51	[1.64-5.38]	< 0.001
Smoking habits (never)	-1.69	[-3.27 to -0.12]	0.03
Hypertension (absence)	-0.75	[-2.72 to 1.22]	0.45
BMI	-0.05	[-0.2 to 0.09]	0.48
Waist circumference	0	[-0.02 to 0.03]	0.75
HbA1c	0.05	[-0.53 to 0.62]	0.87
Diabetes duration	0.07	[-0.01;0.14]	0.09
NDS \geq 6	0.15	[-0.86 to 1.17]	0.76
Ischemic cardiomyopathy	3.73	[2.15-5.32]	< 0.001
eGFR MDRD	-0.06	[-0.01; -0.02]	0.005
hsCRP	-0.16	[-0.47 to 0.16]	0.32
Log IL-6	1.05	[-0.22 to 2.32]	0.10
Triglycerides	0.43	[-0.3 to 1.16]	0.25
Cholesterol total	-0.51	[-1.39 to 0.37]	0.25
HDL	-2.01	[-4.27 to 0.25]	0.08
LDL	0.56	[-1.6 to 0.48]	0.29
Log(sRANKL)	0.04	[-0.39 to 0.46]	0.86
Log(OPG)	1.84	[-1.14 to 4.83]	0.22
Log(OPG/sRANKL)	0	[-0.41 to 0.41]	0.99
iPTH	0.04	[0.01-0.07]	0.01
Log (Corrected calcium)	-9.99	[-26 to 6.02]	0.22
Log(t-ucMGP)	-2.74	[-4.85 to -0.64]	0.01
Log(dp-ucMGP)	1.61	[0.53-2.7]	0.004
Log(FGF-23)	0.56	[-0.03 to 1.16]	0.06
Log(Fetuin A)	-1.82	[-3.36 to -0.29]	0.02
Log(Pentosidine)	2.16	[0.35-3.96]	0.02
Log(Carboxymethyllysine)	1.30	[-2.67 to 5.27]	0.52
Log(MG-H1)	-3.15	[-5.79 to -0.51]	0.02
Log(sRAGE)	0.69	[-0.79 to 2.16]	0.36
Log(IGF-1)	-2.06	[-4.15 to 0.02]	0.05
Standardized IGF-1	-0.24	[-0.75 to 0.27]	0.36
Log adiponectin	1.59	[0.12-3.06]	0.03
TyG INDEX	0.02	[-0.02 to 0.06]	0.25
Statin use	1.62	[-0.93;4.17]	0.21

BMI body mass index, DBP diastolic blood pressure, FGF-23 fibroblast growth factor 23, eGFR MDRD estimated glomerular filtration rate calculated with the modification of diet in renal disease formula, HbA1c haemoglobin A1C, hsCRP high sensitivity C-reactive protein, IGF-1 insulin growth factor-1, IL-6 interleukin 6, iPTH intact parathyroid hormone, MG-H1 Methylglyoxal-derived hydroimidazolone 1, dp-ucMGP desphospho-uncarboxylated matrix gla protein, t-ucMGP Total uncarboxylated matrix gla protein, NDS neuropathy disability score, OPG osteoprotegerin, sRAGE soluble form of receptor for advanced glycation end products, sRANKL soluble form of Receptor Activator of Nuclear factor kB Ligand, TyG triglyceride glucose index

p-values indicating significance are shown in italics type

Table 4 Final multivariate linear regression model: variables independently associated with the progression of calcification score during follow-up (n = 163 patients; mean follow up duration = 31.2 ± 3.86 months)

	β	IC95	p value
Log(calcium score at baseline)	1.02	[1-1.04]	< 0.001
Follow-up duration	0.04	[0.01; 0.06]	0.004
Ischemic cardiomyopathy	0.36	[0.15; 0.57]	0.001
Log(sRANKL)	0.07	[0.02; 0.11]	0.016
Log(OPG/sRANKL)	-0.06	[-0.12-0]	0.03
Triglycerides	0.11	[0.03; 0.2]	0.007
Statin use	0.39	[0.06; 0.72]	0.023

OPG osteoprotegerin, sRANKL soluble form of Receptor Activator of Nuclear factor kB Ligand

p-values indicating significance are shown in italics type

shown that Denosumab attenuates aortic calcification in a murine model [38]. However, in the human, a study has prospectively explored the effects of 12 months of Denosumab treatment on coronary artery calcium scores in 48 patients on hemodialysis [39]. Interestingly, the coronary artery calcium score of patients treated with Denosumab did not increase, even though these patients had very high coronary artery calcium scores at baseline. These data suggest that Denosumab could at least stop the progression of vascular calcification in patients highly exposed to this risk. But interventional studies are needed to determine if targeting RANKL may be beneficial in peripheral arterial calcification progression in type 2 diabetes.

To our knowledge, the effect of fibrates on arterial calcification has not been explored. Hence, targeting triglycerides could be another interesting therapeutic path to take.

As previously reported with accelerated coronary artery calcification in T2D patients with advanced atherosclerosis, statin use is associated with the progression of lower limb arterial calcification in our study [40]. Statins are thought to increase the density of arterial calcification to improve plaque stability but don't seem able to reduce lower limb arterial calcification progression.

The strengths of the present study are its prospective design, the objective quantitative assessment of arterial calcification by CT, the measurement of numerous original markers and the use of a high sensitive kit to measure RANKL. The limitations are the relative small sample size, the absence of a non-diabetic control population, and the absence of distinction between intima and medial calcification [41]. In addition, uncalcified plaques were not assessed in this study.

Conclusions

Lower limb arterial calcification, which has serious clinical consequences, is a frequent pathological process in patients with type 2 diabetes and high cardiovascular risk. It progresses rapidly. The demonstration of the association of RANKL and triglycerides with this progression open up new therapeutic perspectives. Further interventional studies are therefore required to address these issues fully.

Abbreviations

AGEs: Advanced glycation end-products; BMI: Body Mass Index; CML: Carboxymethyllysine; DIACART: Acronym of « DIAbète et Calcification ARterielle »; dp-ucMGP: Desphospho-uncarboxylated Matrix Gla Protein; GFR: Glomerular filtration rate; HbA1c: Glycated hemoglobin; hsCRP: High-sensitivity C-reactive protein; IGF-1: Insulin like growth factor 1; IL-6: Interleukine-6; iPTH: intact parathyroid hormone; MG-H1: Methylglyoxal-derived hydroimidazolone1; NDS: Neuropathy disability score; OPG: Osteoprotegerin; PAOD: Peripheral arterial occlusive disease; RANK: Receptor Activator of Nuclear factor κ B; RANKL: Receptor Activator of Nuclear factor κ B Ligand; sRAGE: Soluble receptor of advanced glycation end-products; sRANKL: Soluble RANKL; t-ucMGP: Total-uncarboxylated Matrix Gla Protein; TyG index: Triglycerides glucose index.

Acknowledgements

The authors thank all the patients, investigators, and CIC-1901 staff members involved in the conduct of the trial. We would also like to thank Dr. Froogh Darakhshan - Hajduch (*Anglais Pour Vous*, Melun, France) for professional editing of the article.

Authors' contributions

FP, C-EA, AC, MH, J-ES, CF-B, SK, AR, PC, J-MD, ZM, RM, DB-R, PG, SJ, CV, J-ML, SL and CA contributed to the acquisition and interpretation of the data. MHD and DH contributed to the acquisition of the data and performed statistical analysis. AH and OB contributed to the study design, to the acquisition and interpretation of the data. OB drafted the report. All authors contributed to the review of the report and approved the final version for submission. OB is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. All authors read and approved the final manuscript.

Funding

DIACART study was funded by the French Ministry of Health grant (PHRC Code projet: P111105; N° ID RCB: 2013-A00896-39). This study was supported by a fund from the Lilly Company. The company was neither involved in the design of the study nor in data collection. The research activities of C.E. Aubert were supported by a doctoral research scholarship from the University of Lausanne. The authors have no relevant conflicts of interest to disclose.

Availability of data and materials

Data are available on request from the corresponding author.

Ethics approval and consent to participate

The study was approved by the local ethics committee (PARIS VI CPP) and registered in ClinicalTrials.gov (Identifier: NCT02431234). All patients were informed about the study objectives and procedure. Participants gave their written informed consent to participation prior to inclusion.

Consent for publication

Not applicable.

Competing interests

No conflict of interest in the area of this study. DIACART study was supported by a fund from the Lilly Company. The company was neither involved in the design of the study nor in data collection.

Author details

¹ Sorbonne Université, Paris, France. ² Assistance Publique-Hôpitaux de Paris (APHP), Diabetology Department, La Pitié Salpêtrière-Charles Foix University Hospital, Paris, France. ³ Unité de Recherche Clinique Salpêtrière - Charles Foix, AP-HP, Hôpitaux Universitaires Pitié Salpêtrière - Charles Foix, 75013 Paris, France. ⁴ Department of Pharmacology and CIC-1421, AP-HP La Pitié Salpêtrière Charles Foix University Hospital, Paris, France. ⁵ INSERM, CIC-1901, Paris, France. ⁶ Institute of Cardiometabolism and Nutrition ICAN, Paris, France. ⁷ Laboratoire d'Imagerie Biomédicale INSERM_1146, CNRS_7371, Paris, France. ⁸ Assistance Publique-Hôpitaux de Paris (APHP), Department of Radiology, La Pitié Salpêtrière-Charles Foix University Hospital, Paris, France. ⁹ Division of Nephrology, Ambroise Paré Hospital, AP-HP, Université Paris-Saclay, Paris, France. ¹⁰ INSERM_1088, Centre Universitaire de Recherche en Santé, Université de Picardie Jules Verne, Amiens, France. ¹¹ Department of Metabolic Biochemistry, La Pitié Salpêtrière-Charles Foix University Hospital (AP-HP), Paris, France. ¹² UTCBS, CNRS UMR8258 - INSERM_1267, Faculty of Pharmacy of Paris, University of Paris, Paris, France. ¹³ University of Reims-Champagne-Ardenne, CNRS, MEDyC UMR 7369, Reims, France. ¹⁴ Laboratory of Biochemistry-Pharmacology-Toxicology, University Hospital of Reims, Maison Blanche Hospital, Reims, France. ¹⁵ Cardiovascular Research Institute CARIM, Maastricht University, Maastricht, The Netherlands. ¹⁶ Department of Endocrine and Oncologic Biochemistry, AP-HP, Pitié-Salpêtrière Hospital, Paris, France. ¹⁷ INSERM U1166, Paris, France. ¹⁸ INSERM, UMR_S 1138, Centre de Recherche des Cordeliers, Paris 06, France. ¹⁹ Département de Santé, Centre de Pharmacopépidémiologie (Cephepi), CIC-1421, Sorbonne Université, INSERM, Institut Pierre Louis d'Epidémiologie et de Santé Publique, AP-HP, Hôpitaux Universitaires Pitié Salpêtrière-Charles Foix, 75013 Paris, France. ²⁰ Sorbonne University, ACTION Study Group, INSERM, UMRS 1166, Institut de Cardiologie, Hôpital Pitié-Salpêtrière (AP-HP), Paris, France. ²¹ Diabetology Department, Pitié-Salpêtrière Hospital, 47-83 Boulevard de l'Hôpital, Paris, France.

Received: 17 May 2020 Accepted: 12 September 2020

Published online: 18 September 2020

References

- Johnson RC, Leopold JA, Loscalzo J. Vascular calcification: pathobiological mechanisms and clinical implications. *Circ Res*. 2006;99:1044–59.
- Toussaint ND, Kerr PG. Vascular calcification and arterial stiffness in chronic kidney disease: implications and management. *Nephrology*. 2007;12:500–9.
- Stabley JN, Towler DA. Arterial calcification in diabetes mellitus: preclinical models and translational implications. *Arterioscler Thromb Vasc Biol*. 2017;37:205–17.
- Bourron O, Aubert CE, Liabeuf S, Cluzel P, Lajat-Kiss F, Dadon M, Komajda M, Mentaverri R, Brazier M, Pierucci A, et al. Below-knee arterial calcification in type 2 diabetes: association with receptor activator of nuclear factor kappaB ligand, osteoprotegerin, and neuropathy. *J Clin Endocrinol Metab*. 2014;99:4250–8.
- Everhart JE, Pettitt DJ, Knowler WC, Rose FA, Bennett PH. Medial arterial calcification and its association with mortality and complications of diabetes. *Diabetologia*. 1988;31:16–23.
- Guzman RJ, Brinkley DM, Schumacher PM, Donahue RM, Beavers H, Qin X. Tibial artery calcification as a marker of amputation risk in patients with peripheral arterial disease. *J Am Coll Cardiol*. 2008;51:1967–74.
- Alessandrino G, Chevalier B, Lefevre T, Sanguineti F, Garot P, Untersee T, Hovasse T, Morice MC, Louvard Y. A clinical and angiographic scoring system to predict the probability of successful first-attempt percutaneous coronary intervention in patients with total chronic coronary occlusion. *JACC Cardiovasc Interv*. 2015;8:1540–8.
- Mattesini A, Di Mario C. Calcium: a predictor of interventional treatment failure across all fields of cardiovascular medicine. *Int J Cardiol*. 2017;231:97–8.
- Evrard S, Delanaye P, Kamel S, Cristol JP, Cavalier E. Vascular calcification: from pathophysiology to biomarkers. *Clin Chim Acta*. 2015;438:401–14.
- Mary A, Hartemann A, Brazier M, Aubert CE, Kamel S, Salem JE, Cluzel P, Liabeuf S, Massy Z, Mentaverri R, et al. Higher parathyroid hormone levels are associated with increased below-the-knee arterial calcification in type 2 diabetes. *Diabetes Metab*. 2018;44:305–8.

11. Harper E, Forde H, Davenport C, Rochfort KD, Smith D, Cummins PM. Vascular calcification in type-2 diabetes and cardiovascular disease: integrative roles for OPG, RANKL and TRAIL. *Vascul Pharmacol*. 2016;82:30–40.
12. Ndip A, Williams A, Jude EB, Serracino-Inglott F, Richardson S, Smyth JV, Boulton AJ, Alexander MY. The RANKL/RANK/OPG signaling pathway mediates medial arterial calcification in diabetic Charcot neuroarthropathy. *Diabetes*. 2011;60:2187–96.
13. Aubert CE, Liabeuf S, Amouyal C, Kemel S, Lajat-Kiss F, Lacorte JM, Halbron M, Carlier A, Salem JE, Funck-Brentano C, et al. Serum concentration and vascular expression of adiponectin are differentially associated with the diabetic calcifying peripheral arteriopathy. *Diabetol Metab Syndr*. 2019;11:32.
14. Rattazzi M, Bertacco E, Puato M, Faggini E, Pualetto P. Hypertension and vascular calcification: a vicious cycle? *J Hypertens*. 2012;30:1885–93.
15. Liabeuf S, Bourron O, Vermeer C, Theuwissen E, Magdeleyns E, Aubert CE, Brazier M, Mentaverri R, Hartemann A, Massy ZA. Vascular calcification in patients with type 2 diabetes: the involvement of matrix Gla protein. *Cardiovasc Diabetol*. 2014;13:85.
16. Abbott CA, Carrington AL, Ashe H, Bath S, Every LC, Griffiths J, Hann AW, Hussein A, Jackson N, Johnson KE, et al. The North-West Diabetes Foot Care Study: incidence of, and risk factors for, new diabetic foot ulceration in a community-based patient cohort. *Diabet Med*. 2002;19:377–84.
17. Schurgers LJ, Teunissen KJ, Knapen MH, Kwaijtaal M, van Diest R, Appels A, Reutelingsperger CP, Cleutjens JP, Vermeer C. Novel conformation-specific antibodies against matrix gamma-carboxyglutamic acid (Gla) protein: undercarboxylated matrix Gla protein as marker for vascular calcification. *Arterioscler Thromb Vasc Biol*. 2005;25:1629–33.
18. Cranenburg EC, Koos R, Schurgers LJ, Magdeleyns EJ, Schoonbrood TH, Landewe RB, Brandenburg VM, Bekers O, Vermeer C. Characterisation and potential diagnostic value of circulating matrix Gla protein (MGP) species. *Thromb Haemost*. 2010;104:811–22.
19. Cranenburg EC, Vermeer C, Koos R, Boumans ML, Hackeng TM, Bouwman FG, Kwaijtaal M, Brandenburg VM, Ketteler M, Schurgers LJ. The circulating inactive form of matrix Gla Protein (ucMGP) as a biomarker for cardiovascular calcification. *J Vasc Res*. 2008;45:427–36.
20. Mahmoudi R, Jaisson S, Badr S, Jaidi Y, Bertholon LA, Novella JL, Gillery P. Post-translational modification-derived products are associated with frailty status in elderly subjects. *Clin Chem Lab Med*. 2019;57:1153–61.
21. Bourron O, Le Bouc Y, Berard L, Kotti S, Brunel N, Ritz B, Leclercq F, Tabone X, Drouet E, Mulak G, et al. Impact of age-adjusted insulin-like growth factor 1 on major cardiovascular events after acute myocardial infarction: results from the fast-MI registry. *J Clin Endocrinol Metab*. 2015;100:1879–86.
22. Guerrero-Romero F, Simental-Mendia LE, Gonzalez-Ortiz M, Martinez-Abundis E, Ramos-Zavala MG, Hernandez-Gonzalez SO, Jacques-Camarena O, Rodriguez-Moran M. The product of triglycerides and glucose, a simple measure of insulin sensitivity. Comparison with the euglycemic-hyperinsulinemic clamp. *J Clin Endocrinol Metab*. 2010;95:3347–51.
23. Agatston AS, Janowitz WR, Hildner FJ, Zusmer NR, Viamonte M Jr, Detrano R. Quantification of coronary artery calcium using ultrafast computed tomography. *J Am Coll Cardiol*. 1990;15:827–32.
24. Baud'huin M, Lamoureux F, Duplomb L, Redini F, Heymann D. RANKL, RANK, osteoprotegerin: key partners of osteoimmunology and vascular diseases. *Cell Mol Life Sci*. 2007;64:2334–50.
25. Panizo S, Cardus A, Encinas M, Parisi E, Valcheva P, Lopez-Ongil S, Coll B, Fernandez E, Valdivielso JM. RANKL increases vascular smooth muscle cell calcification through a RANK-BMP4-dependent pathway. *Circ Res*. 2009;104:1041–8.
26. Schneider F, Saulnier PJ, Gand E, Desvergnès M, Lefort N, Thorin E, Thorin-Trescases N, Mohammedi K, Ragot S, Ricco JB, et al. Influence of micro- and macro-vascular disease and Tumor Necrosis Factor Receptor 1 on the level of lower-extremity amputation in patients with type 2 diabetes. *Cardiovasc Diabetol*. 2018;17:81.
27. Gaudio A, Privitera F, Pulvirenti I, Canzonieri E, Rapisarda R, Fiore CE. Relationships between osteoprotegerin, receptor activator of the nuclear factor κB ligand serum levels and carotid intima-media thickness in patients with type 2 diabetes mellitus. *Panminerva Med*. 2014;56:221–5.
28. Callegari A, Coons ML, Ricks JL, Rosenfeld ME, Scatena M. Increased calcification in osteoprotegerin-deficient smooth muscle cells: dependence on receptor activator of NF-κB ligand and interleukin 6. *J Vasc Res*. 2014;51:118–31.
29. Higgins CL, Isbilir S, Basto P, Chen IY, Vaduganathan M, Vaduganathan P, Reardon MJ, Lawrie G, Peterson L, Morrisett JD. Distribution of alkaline phosphatase, osteopontin, RANK ligand and osteoprotegerin in calcified human carotid atheroma. *Protein J*. 2015;34:315–28.
30. Mohammadpour AH, Shamsara J, Nazemi S, Ghadirzadeh S, Shahsavand S, Ramezani M. Evaluation of RANKL/OPG serum concentration ratio as a new biomarker for coronary artery calcification: a pilot study. *Thrombosis*. 2012;2012:306263.
31. Biscetti F, Bonadia N, Santini F, Angelini F, Nardella E, Pitocco D, Santoliquido A, Filippini M, Landolfi R, Flex A. Sortilin levels are associated with peripheral arterial disease in type 2 diabetic subjects. *Cardiovasc Diabetol*. 2019;18:5.
32. Qamar A, Khetarpal SA, Khera AV, Qasim A, Rader DJ, Reilly MP. Plasma apolipoprotein C-III levels, triglycerides, and coronary artery calcification in type 2 diabetics. *Arterioscler Thromb Vasc Biol*. 2015;35:1880–8.
33. Bjornstad P, Maahs DM, Wadwa RP, Pyle L, Rewers M, Eckel RH, Snell-Bergeon JK. Plasma triglycerides predict incident albuminuria and progression of coronary artery calcification in adults with type 1 diabetes: the coronary artery calcification in type 1 diabetes study. *J Clin Lipidol*. 2014;8:576–83.
34. Kageyama A, Matsui H, Ohta M, Sambuichi K, Kawano H, Notsu T, Imada K, Yokoyama T, Kurabayashi M. Palmitic acid induces osteoblastic differentiation in vascular smooth muscle cells through ACSL3 and NF-κB, novel targets of eicosapentaenoic acid. *PLoS ONE*. 2013;8:e68197.
35. Liao L, Zhou Q, Song Y, Wu W, Yu H, Wang S, Chen Y, Ye M, Lu L. Ceramide mediates Ox-LDL-induced human vascular smooth muscle cell calcification via p38 mitogen-activated protein kinase signaling. *PLoS ONE*. 2013;8:e82379.
36. Biscetti F, Nardella E, Bonadia N, Angelini F, Pitocco D, Santoliquido A, Filippini M, Landolfi R, Flex A. Association between plasma omentin-1 levels in type 2 diabetic patients and peripheral artery disease. *Cardiovasc Diabetol*. 2019;18:74.
37. Bone HG, Wagman RB, Brandi ML, Brown JP, Chapurlat R, Cummings SR, Czerwinski E, Fahrleitner-Pammer A, Kendler DL, Lippuner K, et al. 10 years of denosumab treatment in postmenopausal women with osteoporosis: results from the phase 3 randomised FREEDOM trial and open-label extension. *Lancet Diabetes Endocrinol*. 2017;5:513–23.
38. Helas S, Goettsch C, Schoppet M, Zeitz U, Hempel U, Morawietz H, Kostenuik PJ, Erben RG, Hofbauer LC. Inhibition of receptor activator of NF-κB ligand by denosumab attenuates vascular calcium deposition in mice. *Am J Pathol*. 2009;175:473–8.
39. Iseri K, Watanabe M, Yoshikawa H, Mitsui H, Endo T, Yamamoto Y, Iyoda M, Ryu K, Inaba T, Shibata T. Effects of Denosumab and Alendronate on Bone Health and Vascular Function in Hemodialysis Patients: a Randomized, Controlled Trial. *J Bone Miner Res*. 2019;34:1014–24.
40. Saremi A, Bahn G, Reaven PD. Progression of vascular calcification is increased with statin use in the Veterans Affairs Diabetes Trial (VADT). *Diabetes Care*. 2012;35:2390–2.
41. Konijn LCD, van Overhagen H, Takx RAP, de Jong PA, Veger HTC, Mali W. CT calcification patterns of peripheral arteries in patients without known peripheral arterial disease. *Eur J Radiol*. 2020;128:108973.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.