

RESEARCH

Open Access



SGLT1 and SGLT2 inhibition, circulating metabolites, and cerebral small vessel disease: a mediation Mendelian Randomization study

Yanchen Lv^{1,2*}, Xin Cheng¹ and Qiang Dong¹

Abstract

Background Sodium-glucose cotransporter 2 (SGLT2) and SGLT1 inhibitors may have additional beneficial metabolic effects on circulating metabolites beyond glucose regulation, which could contribute to a reduction in the burden of cerebral small vessel disease (CSVD). Accordingly, we used Mendelian Randomization (MR) to examine the role of circulating metabolites in mediating SGLT2 and SGLT1 inhibition in CSVD.

Methods Genetic instruments for SGLT1/2 inhibition were identified as genetic variants, which were both associated with the expression of encoding genes of SGLT1/2 inhibitors and glycated hemoglobin A1c (HbA1c) level. A two-sample two-step MR was used to determine the causal effects of SGLT1/2 inhibition on CSVD manifestations and the mediating effects of 1400 circulating metabolites linking SGLT1/2 inhibition with CSVD manifestations.

Results A lower risk of deep cerebral microbleeds (CMBs) and small vessel stroke (SVS) was linked to genetically predicted SGLT2 inhibition. Better white matter structure integrity was also achieved, as evidenced by decreased mean diffusivity (MD), axial diffusivity (AD), and radial diffusivity (RD), as well as lower deep (DWMH) and periventricular white matter hyperintensity (PWMH) volume. Inhibiting SGLT2 could also lessen the incidence of severe enlarged perivascular spaces (EPVS) located at white matter, basal ganglia (BG) and hippocampus (HIP). SGLT1 inhibition could preserve white matter integrity, shown as decreased MD of white matter and DWMH volume. The effect of SGLT2 inhibition on SVS and MD of white matter through the concentration of 4-acetamidobutanoate and the cholesterol to oleoyl-linoleoyl-glycerol (18:1 to 18:2) ratio, with a mediated proportion of 30.3% and 35.5% of the total effect, respectively.

Conclusions SGLT2 and SGLT1 inhibition play protective roles in CSVD development. The SGLT2 inhibition could lower the risk of SVS and improve the integrity of white matter microstructure via modulating the level of 4-acetamidobutanoate and cholesterol metabolism. Further mechanistic and clinical studies research are needed to validate our findings.

Keywords Sodium-glucose cotransporter 2 inhibition, Sodium-glucose cotransporter 1 inhibition, Cerebral small vessel disease, Circulating metabolites, Mendelian Randomization

*Correspondence:

Yanchen Lv

lyanchenfudan@163.com

¹Department of Neurology, National Center for Neurological Disorders, National Clinical Research Centre for Aging and Medicine, Huashan Hospital, Fudan University, Shanghai, China

²12 Wulumuqi Zhong Road, 200040 Shanghai, P. R. China



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

Cerebral small vessel disease (CSVD) accounts for nearly 20% of all ischemic strokes and is associated with a high risk of vascular dementia [1, 2]. Type-2 diabetes mellitus (T2DM) is a well-established risk factor for cerebrovascular diseases, such as ischemic stroke [3, 4]. Recent studies have suggested that T2DM is one of the major risk factors for microangiopathy and that patients with diabetes mellitus are more likely to suffer from CSVD [5, 6]. A comprehensive review revealed a substantial correlation between an elevated risk of small subcortical infarcts or white matter hyperintensity (WMH) and prediabetes and increased glycated hemoglobin A1c (HbA1c) level [7]. The Mendelian analysis and longitudinal research pointed out the causal impact of T2DM on CSVD progression [8–11]. Thus, it is crucial to alleviate CSVD in T2DM patients. Since the majority of diabetic patients take drugs to regulate their blood glucose levels, it is important to investigate whether long-term use of anti-diabetic medicines can prevent CSVD.

Sodium-glucose cotransporter 2 (SGLT2) inhibitors are a class of oral anti-diabetic drugs. They can lower serum glucose concentration by inhibiting glucose reabsorption in proximal tubules and promoting urinary glucose excretion [12]. Compelling evidence has shown their beneficial effects beyond glycemic control, but also in improving the prognosis of cardiovascular diseases, heart failure, and kidney diseases [13–16]. In addition, emerging reports have suggested that SGLT2 inhibitors have a neuroprotective effect on critical pathological alterations caused by CSVD, such as the loss of myelin and endothelial integrity [17]. According to recent research, the combination of SGLT1 and SGLT2 inhibitors may significantly reduce stroke risk [18]. The SGLT1 receptor can contribute to neuronal death during ischemia and eliminating it can prevent vascular dementia in a mouse model of CSVD [18, 19]. The above studies indicate that SGLT1/2 inhibitors can prevent CSVD, but the results are controversial, and the underlying metabolic mechanism remains unclear.

Metabolomics is an emerging systems biology technology that can be utilized to explore new biomarkers for disease detection and identify metabolic pathways associated with disease etiology [20]. Prior research has indicated that SGLT2 inhibitors can significantly affect the level of circulating metabolites, especially those related to lipids, amino acids, and ketone bodies [21–23]. Besides, mutations resulting in SGLT1 dysfunction may modify intestinal homeostasis and promote favorable metabolic outcomes [24, 25]. Increasing research has revealed a link between metabolic alternations and CSVD. The large-scale metabolomics study and Mendelian analysis found multiple metabolites associated with imaging markers of CSVD, cognition, and conversion to dementia [26–28].

Accordingly, there may exist metabolic pathways that mediate the impact of SGLT1/2 inhibitors on CSVD progression. However, there is a lack of large-scale longitudinal studies that can validate the causal relationship between the use of SGLT1/2 inhibitors and CSVD manifestations. Moreover, the role of circulating metabolites in the above association can only be partially verified due to the complexity of both the metabolomics and CSVD.

Mendelian Randomization (MR) can use related genetic variants as instrumental variables to examine causal associations, and it has been widely used to unveil the pathophysiological process of diseases in recent years [29]. In the present study, we first performed a two-sample MR study to examine the association between genetic proxies for SGLT2 and SGLT1 inhibitors and CSVD markers. Next, we applied a two-step MR analysis to establish the possible metabolic pathway from SGLT2 inhibition to CSVD using data on circulating metabolites. Our goal was to elucidate the metabolic changes that linked SGLT1/2 inhibition to CSVD.

Methods

Study design

Figure 1 illustrates the study design. The mutual causation between SGLT1/2 inhibition and various CSVD manifestation (Fig. 1A) was assessed using a two-sample MR study. We further conducted a mediation analysis with a two-step MR design to explore whether circulating metabolites could mediate the causal pathway from SGLT1/2 inhibition to CSVD (Fig. 1B). This study followed the Strengthening the Reporting of Observational Studies in Epidemiology Using Mendelian Randomization (STROBE-MR) guideline [30].

Genetic instruments for SGLT2 and SGLT1 inhibitors

According to the previously reported method, genetic variants for drug targets of SGLT1/2 inhibitors were selected using the summary data from the genome-wide association studies (GWAS) involving 344,182 non-diabetic individuals of European ancestry in the UK Biobank [31, 32]. To put it briefly, drug targets and encoding genes of SGLT1/2 inhibitors were identified. Genetic variations close to each gene that were both linked to the expression level of the relevant gene and the glycemic biomarker HbA1c were chosen. Following validation, each target's genetic predictors were generated, with effects quantified as the HbA1c-lowering effect of the target.

Genetic instruments for circulating metabolites

We utilized the most up-to-date and comprehensive GWAS datasets currently available for the human metabolome (Additional file 1) [33]. Based on the Canadian Longitudinal Study on Aging (CLSA) cohort, researchers analyzed data on 1,091 blood metabolites and 309

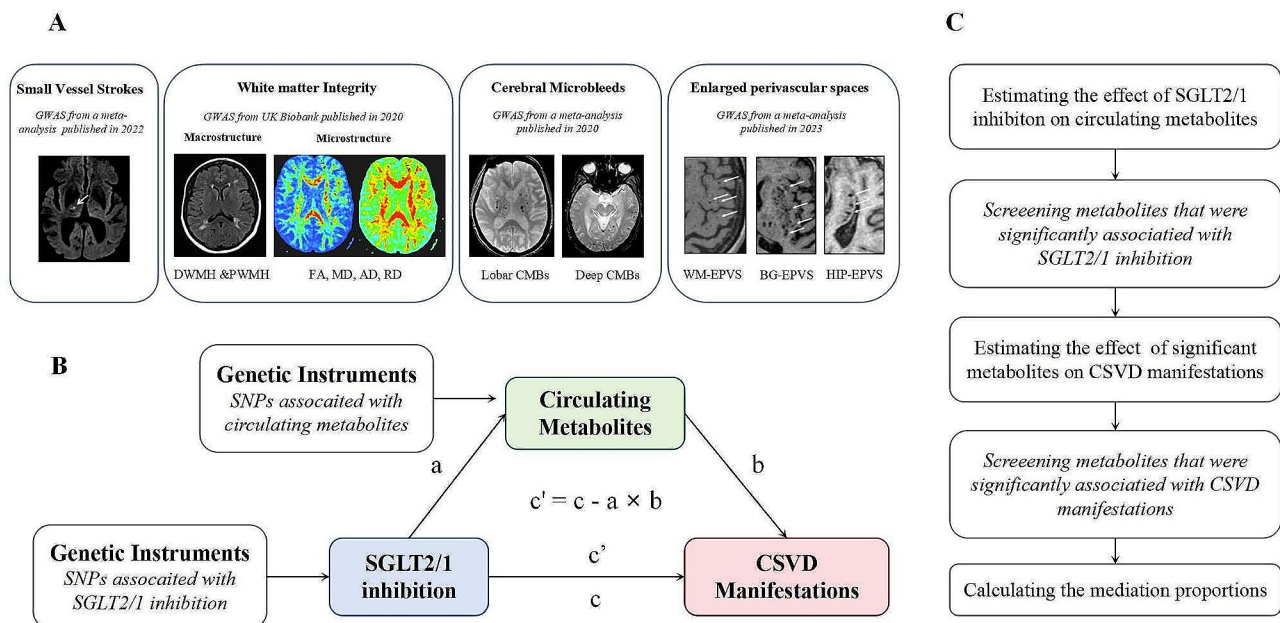


Fig. 1 Overview of the study design. (A) The description of CSVD manifestations used as outcomes. (B) The framework of the two-step MR. (C) The flow diagram of conducting the two-step MR step by step, which involved the selection of circulating metabolites. *SNP, single nucleotide polymorphism; GWAS: Genome-Wide Association Studies; D/PWMH: deep/periventricular white matter hyperintensity; CMBs: cerebral microbleeds; (WM/BG/HIP)-EPVS: (white matter/basal ganglia/hippocampus)-enlarged perivascular spaces; SGLT: sodium-glucose cotransporter; CSVD: cerebral small vessel disease; FA: fractional anisotropy; MD: mean diffusivity; AD: axial diffusivity; RD: radial diffusivity

metabolite ratios by examining a total of 8,299 participants and approximately 15.4 million SNPs. The full GWAS summary statistics of the 1400 biomarkers were publicly available.

CSVD manifestations

To evaluate the causal effect on the risk of small vessel stroke (SVS), we used the data from recently published cross-ancestry GWAS meta-analyses from the GIGAS-STROKE consortium [34]. As one of the clinically overt manifestations of CSVD and a subtype of ischemic stroke, SVS was identified through clinical evaluation and radiological confirmation [34]. SVS was discovered in 13,620 of 1,241,619 participants. The integrity of the white matter macrostructure was assessed using GWAS data of periventricular white matter hyperintensity (PWMH) and deep white matter hyperintensity (DWMH) volume, which were derived from the UK Biobank [35]. To reflect the integrity of white matter microstructure, GWAS data on average fractional anisotropy (FA), mean diffusivity (MD), axial diffusivity (AD), and radial diffusivity (RD) across 21 main white matter tracts were employed [36]. The statistical data on cerebral microbleeds (CMBs) were obtained from a meta-analysis of GWASes involving 11 population-based cohort studies and three case-control or case-only stroke research [37]. CMBs were detected in 3,172 of the 24,354 participants, of which 1,932 were lobar and 1,240 were deep. Up to 40,095 participants from 18 population-based cohorts in genome-wide

association studies provided data on the burden of enlarged perivascular spaces (EPVS). The extensive EPVS burden was observed in the white matter (WM), basal ganglia (BG), and hippocampal (HIP) regions in 9,607 out of 39,822 participants, 9,189 out of 40,000 participants, and 9,339 out of 40,095 participants, respectively [38]. To minimize the risk of population stratification, we chose datasets that included nearly all European ancestry. The details could be found in Additional file 1.

Selection of genetic instruments

We included single nucleotide polymorphisms (SNPs) with a genome-wide significant ($P < 1 \times 10^{-5}$). Then, these SNPs were clumped with a clumping window of 10,000 kb and a linkage disequilibrium (LD) level ($r^2 < 0.001$). Estimated levels of LD from the 1000 Genomes Project based on European samples [39]. Palindromic and ambiguous SNPs were eliminated [40]. The strength of the instruments was judged by F statistics. We removed weak instrumental variables (F-statistics < 10) to avoid bias in MR analysis [41].

Statistical analysis

Effect of SGLT1/2 inhibition on CSVD markers by the MR analysis

We used two-sample MR analyses to explore the causal relationships between SGLT1/2 inhibition and CSVD. The inverse-variance weighted (IVW) regression with a fixed effects model was selected as the primary method

for causal inference [42]. Then, MR-Egger, weighted-median, weighted mode and simple mode methods were conducted to complement and enhance the reliability of the results. The Wald ratio for MR analysis was used when only one genetic instrument was available.

Mediation MR analysis linking SGLT1/2 inhibition with CSVD via circulating metabolites

We further performed a mediation analysis using a two-step MR design to explore whether the level of certain metabolites could mediate the causal pathway from SGLT1/2 inhibition to respective manifestation of CSVD. We first estimated the effect of SGLT1/2 inhibition on circulating metabolites (a in Fig. 1B) using two-sample MR. Second, we evaluated the effect of those metabolites that showed statistically significant associations with SGLT1/2 inhibition on CSVD using two-sample MR (b in Fig. 1B). The total effect gained in previous MR analysis (c in Fig. 1B) can be decomposed into an indirect effect (through mediators, $a \times b$ in Fig. 1B) and a direct effect (without mediators, c' in Fig. 1B) effect [43]. By dividing the indirect effect by the total effect, we could determine the percentage mediated by the mediating effect. Concurrently, the delta approach was utilized to compute 95% confidence intervals (CIs).

Sensitivity analysis

With the aid of funnel plots and Cochran's Q statistic, the heterogeneity between SNPs was evaluated [44]. The

MR-PRESSO [45] and MR-Egger intercept [46] methods were used to identify horizontal pleiotropy. We eliminated any outliers that we found and reassessed the MR causal estimations. A random effects model, which is more resilient to weaker SNP exposure associations was used to evaluate the stability of the results if heterogeneity persisted after elimination of outliers. Ultimately, the impact of each SNP on the total causal estimates was verified using the leave-one-out analysis.

Results

Effect of SGLT2 and SGLT1 inhibition on CSVD

As shown in Additional file 2, ten distinct SNPs were chosen to serve as genetic tools for SGLT2 inhibition, and the F statistics for each SNP were more than 16. rs17683430 (F statistic 59), was used to instrument SGLT1 inhibition. As shown in Fig. 2 and Additional file 3, we found that SGLT2 inhibition was associated with a reduced risk of SVS [odds ratio (OR)=0.161; 95% CI, 0.055–0.466, $P<0.001$], deep CMBs (OR=0.026; 95% CI, 0.002–0.676, $P=0.026$) and better integrity of white matter structure, manifested as lower DWMH volume (Beta = -1.259; 95% CI, -2.041–0.477, $P=0.002$), lower PWMH volume (Beta=0.749; 95% CI, -1.420–0.078, $P=0.029$), decreased RD (Beta = -0.589; 95% CI, -0.940–0.238, $P=0.001$), decreased MD (Beta = -0.697; 95% CI, -1.050–0.344, $P=1.09 \times 10^{-4}$), and reduced AD (Beta = -0.736; 95% CI, -1.097–0.375, $P=6.53 \times 10^{-5}$). SGLT2 inhibition could further reduce the occurrence

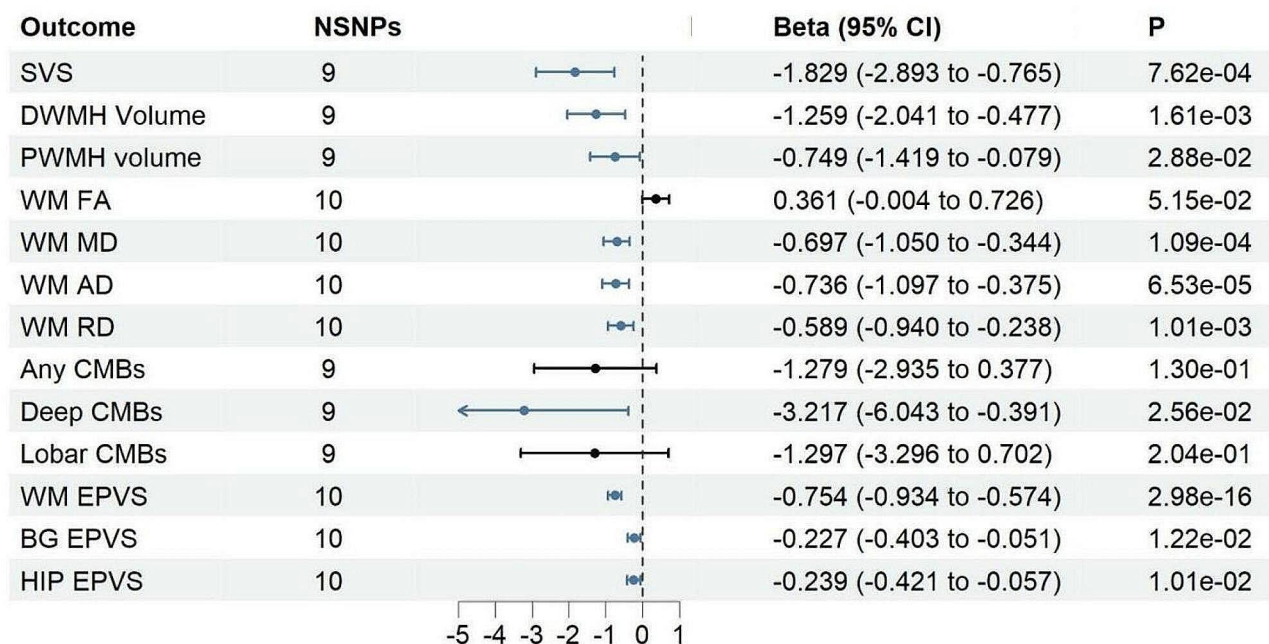


Fig. 2 MR estimates (based on IVW) of the effect of SGLT2 inhibition on CSVD manifestations. *IVW: inverse-variance weighted; MR: Mendelian Randomization; SVS: small vessel stroke; SNP, single nucleotide polymorphism; D/PWMH: deep/periventricular white matter hyperintensity; CMBs: cerebral microbleeds; (WM/BG/HIP)-EPVS: (white matter/basal ganglia/hippocampus)-enlarged perivascular spaces; SGLT: sodium-glucose cotransporter; CSVD: cerebral small vessel disease; FA: fractional anisotropy; MD: mean diffusivity; AD: axial diffusivity; RD: radial diffusivity

of severe EPVS located at white matter (OR=0.470; 95% CI, 0.393–0.564, $P=2.98 \times 10^{-16}$), BG (OR=0.797; 95% CI, 0.668–0.952, $P=0.012$) and HIP regions (OR=0.787, 95% CI, 0.656–0.945, $P=0.010$). SGLT1 inhibition could protect white matter integrity, manifested as lower DWMH volume (Beta = -1.172; 95% CI, -3.315–-0.109, $P=0.036$) and decreased MD (Beta = -0.846; 95% CI, -1.693–-0.001, $P=0.049$) (Fig. 3 and Additional file 4). There was no heterogeneity or horizontal pleiotropy between instruments for analyzing the effect of SGLT1/2 inhibition on CSVD manifestations.

Effect of SGLT1/2 inhibition on circulating metabolites and CSVD via the mediation MR analysis

We estimated the effect of SGLT1/2 inhibition on 1400 circulating metabolites and observed that 168 metabolites were significantly associated with SGLT2 inhibition [Bonferroni-corrected $P < 3.57 \times 10^{-5}$ (0.05/1400)] (Additional file 5) and that no metabolite was associated with SGLT1 inhibition. We further evaluated the effect of 168 circulating metabolites that were significantly associated with SGLT2 inhibition on SVS, deep CMBs, D/PWMH volume, EPVS located at different regions and white matter microstructures, including AD, RD, and MD (Additional file 6). The result showed that seven metabolites were associated with SVS, among which 4-acetamidobutanoate remained a substantial correlation after Bonferroni-correction [$P < 2.98 \times 10^{-4}$ (0.05/168)].

Two metabolites and four metabolites were associated with PWMH volume and DWMH volume respectively, among which 7-methylxanthine had the strongest correlation (PWMH: $P=4.51 \times 10^{-4}$, DWMH: $P=5.82 \times 10^{-3}$), but it did not present a significant relationship after Bonferroni-correction. In terms of white matter microstructure, eight, six and seven metabolites were associated with MD, AD and RD of white matter, respectively. After Bonferroni-correction, the cholesterol to oleoyl-linoleoyl-glycerol (18:1 to 18:2) ratio ($P=1.91 \times 10^{-4}$) was significantly associated with RD of white matter. For deep CMBs, seven metabolites showed a suggestively significant ($P < 0.05$, $P_{\text{adj}} > 0.05$) association. Seven, six and eleven metabolites were associated with BG-EPVS, WM-EPVS and HIP-EPVS respectively, but none showed a statistical association after Bonferroni-correction.

We observed an indirect effect of SGLT2 inhibition on SVS through 4-acetamidobutanoate, with a mediated proportion of 30.3% (95% CI, 13.5–47.1%, $P=4.19 \times 10^{-4}$) of the total effect (Fig. 4A). The indirect effect of SGLT2 inhibition on RD of white matter through the cholesterol to oleoyl-linoleoyl-glycerol (18:1 to 18:2) ratio had a mediated proportion of 35.5% (95% CI, 14.4–56.6%, $P=9.96 \times 10^{-4}$) (Fig. 4B). There was no evidence of heterogeneity and no horizontal pleiotropy among these associations.

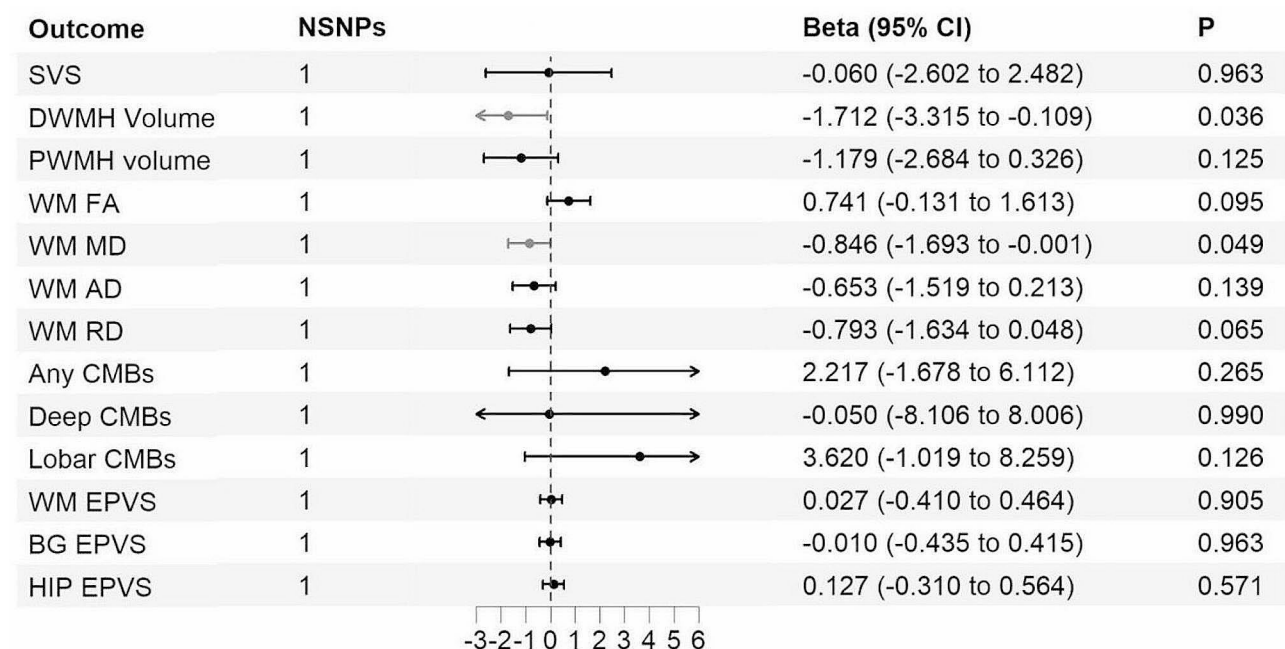
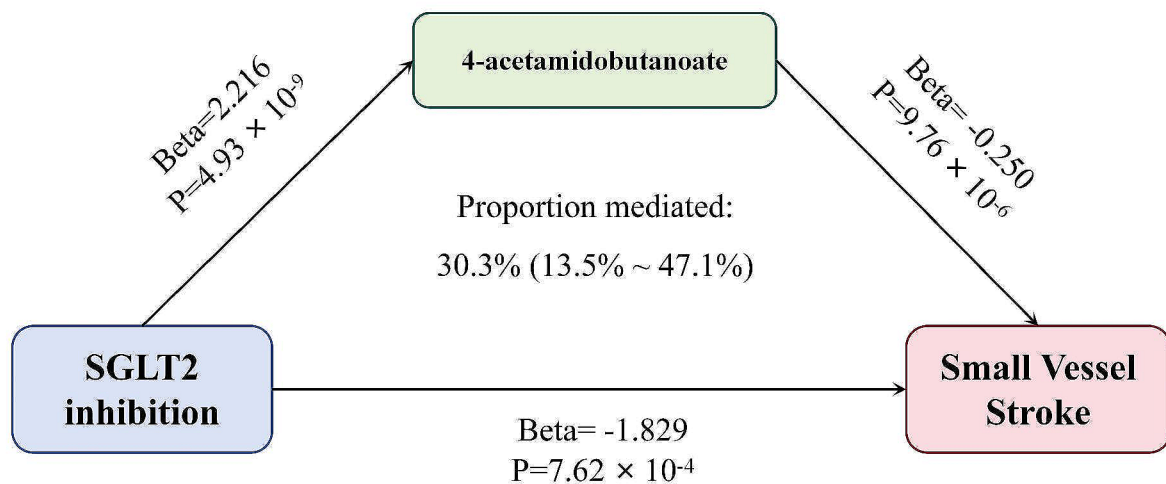


Fig. 3 MR estimates (based on Wald ratio) of the effect of SGLT1 inhibition on CSVD manifestations. *MR: Mendelian Randomization; SVS: small vessel stroke; SNP, single nucleotide polymorphism; D/PWMH: deep/periventricular white matter hyperintensity; CMBs: cerebral microbleeds; (WM/BG/HIP)-EPVS: (white matter/basal ganglia/hippocampus)-enlarged perivascular spaces; SGLT: sodium-glucose cotransporter; CSVD: cerebral small vessel disease; FA: fractional anisotropy; MD: mean diffusivity; AD: axial diffusivity; RD: radial diffusivity

A



B

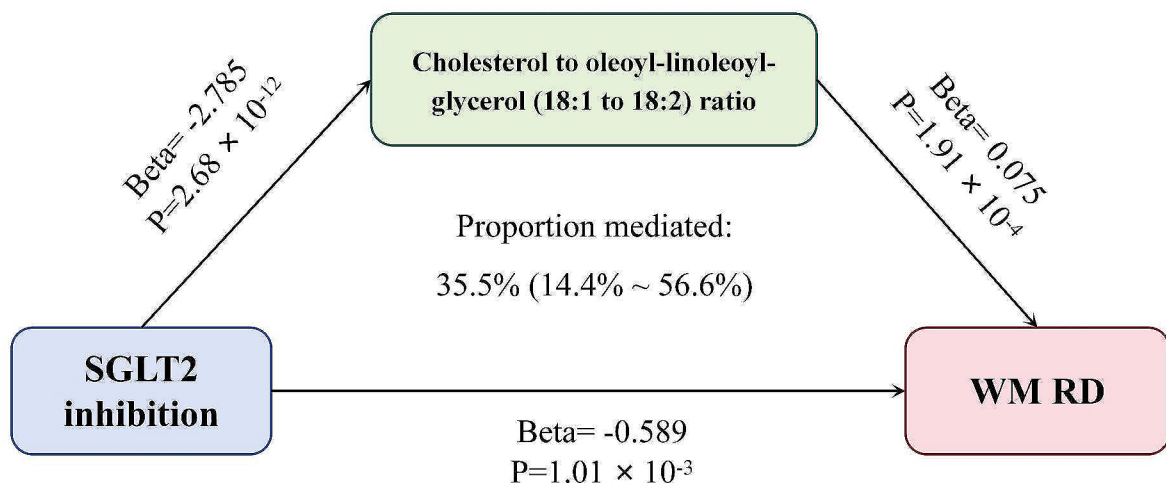


Fig. 4 The potential causal evidence summarized from the MR analysis. (A) The mediation effects of 4-acetamidobutanoate on the association between SGLT2 inhibitors and SVS. (B) The mediation effects of cholesterol to oleoyl-linoleoyl-glycerol (18:1 to 18:2) ratio on the association between SGLT2 inhibitors and the RD of white matter. *MR: Mendelian Randomization; SVS: small vessel stroke; SNP, single nucleotide polymorphism; WM: white matter; SGLT: sodium-glucose cotransporter; RD: radial diffusivity

Discussion

Principal findings

In the present study, we evaluated the associations between genetically predicted SGLT1/2 inhibition and CSVD and investigated the mediating role of circulating metabolites in the relationship. Our study indicated that the genetic variation for targets of SGLT2 inhibition was associated with a lower risk of SVS, deep CMBs and a lower burden of EPVS. Both SGLT2 and SGLT1 inhibition might lead to better integrity of white matter structure. The total concentration of 4-acetamidobutanoate and the cholesterol to oleoyl-linoleoyl-glycerol (18:1 to

18:2) ratio could partially mediate the effect of SGLT2 inhibition on SVS and RD of white matter, respectively.

Association between SGLT1/2 inhibition and CSVD

Among studies investigating the effect of SGLT1/2 inhibition on cerebrovascular dysfunction, most research focused on the risk of stroke. Large clinical trials and meta-analyses have investigated the role of SGLT2 inhibition in fatal or non-fatal stroke and ischemic stroke. Intracerebral ventricular phlorizin [47, 48] and anti-sense SGLT1 mRNA treatment [49] have been indicated to decrease the size of the infarct. Additionally, studies demonstrated a relatively stronger protective effect of

SGLT1/SGLT2 inhibition against stroke [50]. However, no study revealed the effects of SGLT1/SGLT2 inhibition on CSVD progression such as the occurrence of SVS and increased volume of WMH. The present study first reported the potential protective effect of SGLT2 and SGLT1 inhibition on CSVD. In line with our findings, numerous prior studies have demonstrated the preventive effect of SGLT2 and SGLT1 inhibitors on the pathological alterations linked to CSVD. The primary expression site of SGLT2 in the brain is microvasculature [51], which is closely related to the neurovascular unit (NVU), a fundamental biological system. As a key structural and functional component of the blood–brain barrier (BBB), NVU plays an important role in CSVD pathogenesis [52]. The NVU's cell and myelin ultrastructural remodeling, including endothelial cells (ECs) and cortical matter aberration as well as the attenuation or loss of EC tight and adherent junctions of the BBB, could be avoided by suppressing SGLT2 expression with empagliflozin (EMP) [53]. Additionally, much evidence has suggested neuroinflammation as one of the main pathophysiological mechanisms of CSVD progression, with microglia serving as the catalyst for this process [54, 55]. Following EMP treatment, there was a decrease in the levels of tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6), and monocyte chemoattractant protein-1 (MCP-1)—all of which were strongly linked to M1 microglia activation, which had a high capacity for phagocytic activity [56–58]. Hierro-Bujalance et al. also observed reduced microglia burden in the parenchyma of db/db and APP/PS1xdb/db mice treated with EMP [59]. The reduction of oxidative stress, as seen by lower levels of malondialdehyde, higher catalase activity, and higher glutathione, may also underly the neuroprotective effect of empagliflozin [60]. Like SGLT2, cerebral SGLT1 exists in neurons and ECs of small vessels [61, 62]. Under hypoxic conditions, ECs from small arteries express more SGLT1 [62, 63], which triggers microglia activation and causes vascular cognitive impairments in CSVD [19]. It also enhances the expression of MCP-1, IL-1 β , TNF- α , and IL-6 in neurons and/or ECs [19]. Thus, SGLT1 gene deletion may protect against CSVD by inhibiting macrophage infiltration and microglia activation.

Association between SGLT1/2 inhibition and circulating metabolites

The effects of SGLT2 inhibitors on metabolites have been frequently reported. Our results further emphasized the impact of SGLT2 inhibitors on the metabolism of amino acids, especially leucine and isoleucine, both of which belonged to branched-chain amino acids (BCAAs). Newgard et al. has reported the link between elevated plasma concentrations of BCAA and insulin resistance [64]. However, a recent study found that 20 g BCAA daily for

four weeks could improve glucose metabolism [65]. Furthermore, an investigation revealed that an eight-week intake of BCAAs reduced liver fat accumulation [66]. Furuya et al. also detected higher plasma levels of valine and leucine after one-week administration of SGLT2 inhibitor [67]. Together with our results, these findings implied that BCAA improved insulin resistance rather than worsened it and might regulate the ameliorating effect of SGLT2 inhibitors. SGLT1 inhibitors were less likely to affect metabolite levels compared with SGLT2 inhibitors. Prior research has suggested that the positive effects of SGLT1 inhibitors may be attributed to that they can regulate inflammatory cytokines to prevent macrophage infiltration and microglia activation [19]. Consequently, the effect of SGLT1 inhibitors could not be fully captured by the metabolites included in the present investigation. The exact mechanism by which SGLT1 inhibitors preserved the integrity of white matter should be further investigated.

Mediating role of circulating metabolites in the association between SGLT2 inhibition and CSVD manifestations

It was noteworthy that genetically predicted SGLT2 inhibition might lower the risk of SVS by modulating the level of 4-acetamidobutanoate. As a urea cycle product, 4-acetamidobutanoate was associated with the polyamine (PA) metabolism. The PA was secreted from intracellular compartments in various cerebral nervous system (CNS) traumas, such as focal cerebral ischemia in the ischemic cascade [68, 69]. In rat brain homogenates, it acted as free radical scavengers to lessen lipid peroxidation brought on by prooxidant agents like quinolinic acid, iron (Fe²⁺), and sodium nitroprusside [70, 71]. Due to the capacity of PA to raise nitric oxide bioavailability, lower oxidative stress, change structural variables, and promote autophagy, it might improve EC function [72]. 4-acetamidobutanoate was also reported to be a derivative of gamma-aminobutyric acid (GABA) [73], a crucial inhibitory neurotransmitter in the human CNS. The efficacy of hyperpolarization and chloride transport across the postsynaptic membrane could be enhanced by GABA neurotransmission. As demonstrated by the neuroprotection of GABA receptor agonists in animal stroke models, these actions could balance the harmful effects of glutamate during cerebral ischemia. GABA levels in the cerebro-spinal fluid (CSF) and plasma were shown to be lower in patients suffering from global cerebral ischemia and acute ischemic stroke [74].

The mediating role of the cholesterol to oleoyl-linoleoyl-glycerol (18:1 to 18:2) ratio in the connection between SGLT2 inhibition and white matter microstructure further highlighted the importance of cholesterol and its homeostasis in CSVD. Cholesterol played a pivotal role in the brain and brain diseases. The high

cholesterol diet-induced hypercholesterolemia could cause microgliosis, astrogliosis and white matter inflammation [75]. Animal models of familial hypercholesterolemia also exhibited angiopathology associated with white matter injuries like BBB disruption [76, 77]. Numerous mechanisms, including intraluminal cell accumulations [78], reduced vasodilatory response [79], severe inflammation, and thrombosis [80, 81], connected hypercholesterolemia to the pathological alterations of small arteries and capillaries in CSVD. In addition to total cholesterol levels, the integrity of white matter has been shown to destroy cholesterol homeostasis, manifested as elevated low-density lipoprotein cholesterol (LDL-c) level and decreased high-density lipoprotein cholesterol (HDL-c) concentration [82]. Nevertheless, LDL-c, particularly small and dense LDL-c (sdLDL-c), was easily oxidized and slowly cleared, thus entering the artery wall to cause atherosclerosis [83], which was harmful to white matter, HDL-c maintains the integrity of white matter by regulating the function of vascular smooth muscle cells, improving EC function, and maintaining BBB integrity [84, 85]. Oleoyl-linoleoyl-glycerol (18:1 to 18:2) [DAG (36:3)] is a type of diacylglycerol. By stimulating the production and release of HDL-c, diacylglycerol was able to raise HDL-c levels, while at the same time lowering total cholesterol and LDL-c levels by speeding up the elimination of LDL-c and blocking the synthesis of cholesterol. Increased plasma levels of DAG (36:3) were associated with moderate to severe obstructive sleep apnea (OSA) [86], which was an independent risk factor for asymptomatic CSVD, particularly WMH and acute small subcortical infarcts [87].

Strengths and limitations

As far as we know, this study stands out as the first to delve into the connection between SGLT2 and SGLT1 inhibition, circulating metabolites, and CSVD manifestations using MR analysis. Moreover, the genetic evidence supporting the possible mechanism of SGLT2 inhibition in preventing CSVD was presented. Still, some limitations merit consideration. To begin with, genetic variations that mimic SGLT2 inhibition revealed the lifelong effects of SGLT2 inhibitors; however, these effects may differ from that of SGLT2 inhibitors used for a short period. Besides, given that CSVD is an age-related and progressive disease, patients in various age groups or disease stages might respond differently to SGLT inhibition. As such, the direction rather than the amount of the probable causal influence can be better investigated in our study. Experiments and large-scale clinical trials based on different population such as very elderly population are warranted for further investigation. Second, as a complex disease, CSVD can present different types and distribution patterns of imaging markers. Although

the most representative imaging markers were selected, we could not cover all CSVD manifestations. Moreover, there were overlapped samples in GWAS of SGLT1/2 inhibition and CSVD manifestations, which might cause a bias in the potential causal effect estimate in the case of the weak instruments [88]. Nonetheless, the genetic variations for SGLT1/2 inhibition were highly correlated with exposure, as shown by high F-statistics, suggesting the validity of our findings. Finally, because the data used in the present study were derived mainly from the European population, the extrapolation of the results to other ethnic populations should be considered with caution.

Conclusions

In conclusion, our findings indicate that SGLT2 and SGLT1 may prevent CSVD development. The SGLT2 inhibition may lower the risk of SVS and improve the integrity of white matter microstructure by modulating the level of 4-acetamidobutanoate and cholesterol metabolism. These findings provide novel insights into the value of SGLT inhibition in preventing CSVD and might guide future mechanistic and clinical studies.

Abbreviations

AD	Axial diffusivity
BBB	Blood–brain barrier
BCAA	Branched chain amino acids
CI	Confidence intervals
CLSA	Canadian Longitudinal Study on Aging
CMBs	Cerebral microbleeds
CNS	Cerebral nervous system
CSF	Cerebro-spinal fluid
CSVD	Cerebral small vessel disease
DAG	Diacylglycerol
D/PWMH	Deep/periventricular white matter hyperintensity
(WM/BG/HIP)-EPVS	(white matter/basal ganglia/hippocampus)-enlarged perivascular spaces
EC	Endothelial cell
EMP	Empagliflozin
FA	Fractional anisotropy
GABA	Gamma-aminobutyric acid
GWAS	Genome-Wide Association Studies
HbA1C	Glycated hemoglobin level
HDL-c	High-density lipoprotein cholesterol
IL	Interleukin
IWV	Inverse–variance weighted
LD	Linkage disequilibrium
LDL-c	Low-density lipoprotein cholesterol
MCP-1	Monocyte chemoattractant protein-1
MD	Mean diffusivity
MR	Mendelian Randomization
NYU	Neurovascular unit
OR	Odds ratio
OSA	Obstructive sleep apnea
PA	Polyamine
RD	Radial diffusivity/radial diffusivity
sdLDL-C	Small and dense low-density lipoprotein cholesterol
SGLT	Sodium–glucose cotransporter
SNP	Single nucleotide polymorphism
STROBE-MR	Strengthening the Reporting of Observational Studies in Epidemiology Using Mendelian Randomization
SVS	Small vessel stroke
TNF	Tumor necrosis factor
WM	White matter

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12933-024-02255-6>.

Additional file 1: Table S1. Detailed information for genome-wide association study (GWAS) statistics used in the present study. **Additional file 2: Table S2.** Instrumental variables for SGLT2 and SGLT1 inhibition. **Additional file 3: Table S3.** MR estimates of the effect of SGLT2 inhibition on CSVD manifestations. **Additional file 4: Table S4.** MR estimates of the effect of SGLT1 inhibition on CSVD manifestations. **Additional file 5: Table S5.** Significant ($p < 0.05$) MR estimates of the effect of SGLT2 inhibition on 1400 metabolites. **Additional file 6: Table S6.** The effects of SGLT2 inhibition on circulating metabolites and the effects of metabolites on CSVD manifestations.

Acknowledgements

We thank all studies for providing genotype and phenotype data supporting the original work of all GWASes used in this study.

Author contributions

Yanchen Lv contributed to the design of the work, the acquisition, analysis, interpretation of data and have drafted the manuscript; Qiang Dong and Xin Cheng contributed to the substantial revision of the work. All authors read and approved the final manuscript.

Funding

This study was funded by National Natural Science Foundation of China (81971123, 82271352), Shanghai Municipal Committee of Science and Technology (20Z11900802), and Shanghai Municipal Health Commission (2022XD022).

Data availability

All GWAS summary statistics used for the included MR analyses could be found in Supplemental data. R scripts for these analyses could be shared upon request (lvyanchenfudan@163.com).

Declarations

Ethics approval and consent to participate

All studies included in cited genome-wide association studies had been approved by a relevant review board, and participants had provided written informed consent.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 14 February 2024 / Accepted: 29 April 2024

Published online: 07 May 2024

References

1. Iadecola C, Duering M, Hachinski V, Joutel A, Pendlebury ST, Schneider JA, et al. Vascular cognitive impairment and dementia: JACC Scientific Expert Panel. *J Am Coll Cardiol*. 2019;73(25):3326–44.
2. O'Brien JT, Thomas A. Vascular dementia. *Lancet (London England)*. 2015;386(10004):1698–706.
3. Cosentino F, Grant PJ, Aboyans V, Bailey CJ, Ceriello A, Delgado V, et al. 2019 ESC guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD. *Eur Heart J*. 2020;41(2):255–323.
4. Meschia JF, Bushnell C, Boden-Albala B, Braun LT, Bravata DM, Chaturvedi S, et al. Guidelines for the primary prevention of stroke: a statement for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke*. 2014;45(12):3754–832.
5. Park JH, Ryoo S, Kim SJ, Kim GM, Chung CS, Lee KH, et al. Differential risk factors for lacunar stroke depending on the MRI (white and red) subtypes of microangiopathy. *PLoS ONE*. 2012;7(9):e44865.
6. Wang DQ, Wang L, Xia XS, Wei MM, Tian XL, Wang LF, et al. Clinical and MRI features about two types of silent cerebral small-vessel disease in type-2 diabetes mellitus: a retrospective cross-sectional study in a tertiary hospital. *Quant Imaging Med Surg*. 2022;12(4):2385–96.
7. Zhou JB, Tang XY, Han YP, Luo FQ, Cardoso MA, Qi L. Prediabetes and structural brain abnormalities: evidence from observational studies. *Diab/Metab Res Rev*. 2020;36(4):e3261.
8. Liu J, Rutten-Jacobs L, Liu M, Markus HS, Traylor M. Causal impact of type 2 diabetes Mellitus on Cerebral Small Vessel Disease: a mendelian randomization analysis. *Stroke*. 2018;49(6):1325–31.
9. Marseglia A, Fratiglioni L, Kalpouzos G, Wang R, Bäckman L, Xu W. Prediabetes and diabetes accelerate cognitive decline and predict microvascular lesions: a population-based cohort study. *Alzheimer's Dement J Alzheimer's Assoc*. 2019;15(1):25–33.
10. Lucatelli P, Montisci R, Sanfilippo R, Sacconi B, Suri JS, Catalano C, et al. Is there an association between leukoaraiosis volume and diabetes? *J Neuroradiol = J de Neuroradiologie*. 2016;43(4):273–9.
11. Georgakis MK, Harshfield EL, Malik R, Franceschini N, Langenberg C, Wareham NJ, et al. Diabetes Mellitus, glycemic traits, and Cerebrovascular Disease: a mendelian randomization study. *Neurology*. 2021;96(13):e1732–42.
12. Heerspink HJ, Perkins BA, Fitchett DH, Husain M, Cherney DZ. Sodium glucose cotransporter 2 inhibitors in the treatment of diabetes Mellitus: Cardiovascular and kidney effects, potential mechanisms, and clinical applications. *Circulation*. 2016;134(10):752–72.
13. Wiviott SD, Raz I, Bonaca MP, Mosenzon O, Kato ET, Cahn A, et al. Dapagliflozin and Cardiovascular outcomes in type 2 diabetes. *N Engl J Med*. 2019;380(4):347–57.
14. Neal B, Perkovic V, Mahaffey KW, de Zeeuw D, Fulcher G, Erondou N, et al. Canagliflozin and Cardiovascular and renal events in type 2 diabetes. *N Engl J Med*. 2017;377(7):644–57.
15. Cannon CP, Pratley R, Dagogo-Jack S, Mancuso J, Huyck S, Masiukiewicz U, et al. Cardiovascular outcomes with Ertugliflozin in Type 2 diabetes. *N Engl J Med*. 2020;383(15):1425–35.
16. Bhatt DL, Szarek M, Pitt B, Cannon CP, Leiter LA, McGuire DK, et al. Sotagliflozin in patients with diabetes and chronic kidney disease. *N Engl J Med*. 2021;384(2):129–39.
17. Pawlos A, Broncel M, Woźniak E, Gorzelak-Pabiś P. Neuroprotective effect of SGLT2 inhibitors. *Molecules*. 2021;26(23).
18. Pitt B, Steg G, Leiter LA, Bhatt DL. The role of combined SGLT1/SGLT2 inhibition in reducing the incidence of stroke and myocardial infarction in patients with type 2 diabetes Mellitus. *Cardiovasc Drugs Ther*. 2022;36(3):561–7.
19. Ishida N, Saito M, Sato S, Koepsell H, Taira E, Hirose M. SGLT1 participates in the development of vascular cognitive impairment in a mouse model of small vessel disease. *Neurosci Lett*. 2020;727:134929.
20. Clish CB. Metabolomics: an emerging but powerful tool for precision medicine. *Cold Spring Harbor Mol case Stud*. 2015;1(1):a000588.
21. Kappel BA, Lehrke M, Schütt K, Artati A, Adamski J, Lebherz C, et al. Effect of Empagliflozin on the metabolic signature of patients with type 2 diabetes Mellitus and Cardiovascular Disease. *Circulation*. 2017;136(10):969–72.
22. Katano S, Yano T, Kouzu H, Nagaoka R, Numazawa R, Yamano K, et al. Elevated circulating level of β -aminoisobutyric acid (BAIBA) in heart failure patients with type 2 diabetes receiving sodium-glucose cotransporter 2 inhibitors. *Cardiovasc Diabetol*. 2022;21(1):285.
23. Szekeres Z, Toth K, Szabados E. The effects of SGLT2 inhibitors on lipid metabolism. *Metabolites*. 2021;11(2).
24. Lehmann A, Hornby PJ. Intestinal SGLT1 in metabolic health and disease. *Am J Physiol Gastrointest Liver Physiol*. 2016;310(11):G887–98.
25. Seidemann SB, Feofanova E, Yu B, Franceschini N, Claggett B, Kuokkanen M, et al. Genetic variants in SGLT1, glucose tolerance, and cardiometabolic risk. *J Am Coll Cardiol*. 2018;72(15):1763–73.
26. Harshfield EL, Markus HS. Association of Baseline Metabolomic profiles with Incident Stroke and Dementia and with imaging markers of Cerebral Small Vessel Disease. *Neurology*. 2023;101(5):e489–501.
27. Harshfield EL, Sands CJ, Tuladhar AM, de Leeuw FE, Lewis MR, Markus HS. Metabolomic profiling in small vessel disease identifies multiple associations with disease severity. *Brain*. 2022;145(7):2461–71.
28. Sun Y, Guo Y, Li HQ, Tan L, Feng JF, Cheng W, et al. Associations of circulating metabolites with cerebral white matter hyperintensities. *J Neurochem*. 2023;166(2):414–23.

29. Evans DM, Davey Smith G. Mendelian randomization: New Applications in the coming age of hypothesis-free causality. *Annu Rev Genom Hum Genet.* 2015;16:327–50.
30. Skrivankova VW, Richmond RC, Woolf BAR, Yarmolinsky J, Davies NM, Swanson SA, et al. Strengthening the reporting of Observational studies in Epidemiology using mendelian randomization: the STROBE-MR Statement. *JAMA.* 2021;326(16):1614–21.
31. Li J, Yu Y, Sun Y, Yu B, Tan X, Wang B, et al. SGLT2 inhibition, circulating metabolites, and atrial fibrillation: a mendelian randomization study. *Cardiovasc Diabetol.* 2023;22(1):278.
32. Zhao SS, Rajasundaram S, Karhunen V, Alam U, Gill D. Sodium-glucose cotransporter 1 inhibition and gout: mendelian randomisation study. *Semin Arthritis Rheum.* 2022;56:152058.
33. Chen Y, Lu T, Pettersson-Kymmer U, Stewart ID, Butler-Laporte G, Nakanishi T, et al. Genomic atlas of the plasma metabolome prioritizes metabolites implicated in human diseases. *Nat Genet.* 2023;55(1):44–53.
34. Mishra A, Malik R, Hachiya T, Jürgenson T, Namba S, Posner DC, et al. Stroke genetics informs drug discovery and risk prediction across ancestries. *Nature.* 2022;611(7934):115–23.
35. Elliott LT, Sharp K, Alfaro-Almagro F, Shi S, Miller KL, Douaud G, et al. Genome-wide association studies of brain imaging phenotypes in UK Biobank. *Nature.* 2018;562(7726):210–6.
36. Zhao B, Li T, Yang Y, Wang X, Luo T, Shan Y, et al. Common genetic variation influencing human white matter microstructure. *Volume 372.* New York, NY: Science; 2021. 6548.
37. Knol MJ, Lu D, Traylor M, Adams HHH, Romero JRJ, Smith AV, et al. Association of common genetic variants with brain microbleeds: a genome-wide association study. *Neurology.* 2020;95(24):e3331–43.
38. Duperron MG, Knol MJ, Le Grand Q, Evans TE, Mishra A, Tsuchida A, et al. Genomics of perivascular space burden unravels early mechanisms of cerebral small vessel disease. *Nat Med.* 2023;29(4):950–62.
39. Abecasis GR, Altshuler D, Auton A, Brooks LD, Durbin RM, Gibbs RA, et al. A map of human genome variation from population-scale sequencing. *Nature.* 2010;467(7319):1061–73.
40. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D et al. The MR-Base platform supports systematic causal inference across the human phenome. *eLife.* 2018;7.
41. Burgess S, Thompson SG. Avoiding bias from weak instruments in mendelian randomization studies. *Int J Epidemiol.* 2011;40(3):755–64.
42. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol.* 2013;37(7):658–65.
43. Carter AR, Sanderson E, Hammerton G, Richmond RC, Davey Smith G, Heron J, et al. Mendelian randomisation for mediation analysis: current methods and challenges for implementation. *Eur J Epidemiol.* 2021;36(5):465–78.
44. Tan JS, Liu NN, Guo TT, Hu S, Hua L. Genetically predicted obesity and risk of deep vein thrombosis. *Thromb Res.* 2021;207:16–24.
45. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from mendelian randomization between complex traits and diseases. *Nat Genet.* 2018;50(5):693–8.
46. Burgess S, Thompson SG. Interpreting findings from mendelian randomization using the MR-Egger method. *Eur J Epidemiol.* 2017;32(5):377–89.
47. Yamazaki Y, Ogiwara S, Harada S, Tokuyama S. Activation of cerebral sodium-glucose transporter type 1 function mediated by post-ischemic hyperglycemia exacerbates the development of cerebral ischemia. *Neuroscience.* 2015;310:674–85.
48. Yamazaki Y, Harada S, Tokuyama S. Post-ischemic hyperglycemia exacerbates the development of cerebral ischemic neuronal damage through the cerebral sodium-glucose transporter. *Brain Res.* 2012;1489:113–20.
49. Yamazaki Y, Harada S, Wada T, Yoshida S, Tokuyama S. Sodium transport through the cerebral sodium-glucose transporter exacerbates neuron damage during cerebral ischaemia. *J Pharm Pharmacol.* 2016;68(7):922–31.
50. Pitt B, Bhatt DL. Does SGLT1 inhibition add benefit to SGLT2 inhibition in type 2 diabetes? *Circulation.* 2021;144(1):4–6.
51. Nguyen T, Wen S, Gong M, Yuan X, Xu D, Wang C et al. Dapagliflozin activates neurons in the Central Nervous System and regulates Cardiovascular activity by inhibiting SGLT-2 in mice. *Diabetes, metabolic syndrome and obesity: targets and therapy.* 2020;13:2781–99.
52. Yang Q, Wei X, Deng B, Chang Z, Jin D, Huang Y, et al. Cerebral small vessel disease alters neurovascular unit regulation of microcirculation integrity involved in vascular cognitive impairment. *Neurobiol Dis.* 2022;170:105750.
53. Hayden MR, Grant DG, Aroor AR, DeMarco VG. Empagliflozin ameliorates type 2 Diabetes-Induced ultrastructural remodeling of the neurovascular unit and Neuroglia in the female db/db mouse. *Brain Sci.* 2019;9(3).
54. Terasaki Y, Liu Y, Hayakawa K, Pham LD, Lo EH, Ji X, et al. Mechanisms of neurovascular dysfunction in acute ischemic brain. *Curr Med Chem.* 2014;21(18):2035–42.
55. Wardlaw JM, Smith C, Dichgans M. Small vessel disease: mechanisms and clinical implications. *Lancet Neurol.* 2019;18(7):684–96.
56. Han JH, Oh TJ, Lee G, Maeng HJ, Lee DH, Kim KM, et al. The beneficial effects of empagliflozin, an SGLT2 inhibitor, on atherosclerosis in ApoE (-/-) mice fed a western diet. *Diabetologia.* 2017;60(2):364–76.
57. Dimitriadis GK, Nasiri-Ansari N, Agrogiannis G, Kostakis ID, Randeve MS, Nikiteas N, et al. Empagliflozin improves primary haemodynamic parameters and attenuates the development of atherosclerosis in high fat diet fed APOE knockout mice. *Mol Cell Endocrinol.* 2019;494:110487.
58. Ganbaatar B, Fukuda D, Shinohara M, Yagi S, Kusunose K, Yamada H, et al. Empagliflozin ameliorates endothelial dysfunction and suppresses atherogenesis in diabetic apolipoprotein E-deficient mice. *Eur J Pharmacol.* 2020;875:173040.
59. Hierro-Bujalance C, Infante-García C, Del Marco A, Herrera M, Carranza-Naval MJ, Suarez J, et al. Empagliflozin reduces vascular damage and cognitive impairment in a mixed murine model of Alzheimer's disease and type 2 diabetes. *Alzheimers Res Ther.* 2020;12(1):40.
60. Amin EF, Rifaai RA, Abdel-Latif RG. Empagliflozin attenuates transient cerebral ischemia/reperfusion injury in hyperglycemic rats via repressing oxidative-inflammatory-apoptotic pathway. *Fundam Clin Pharmacol.* 2020;34(5):548–58.
61. Poppe R, Karbach U, Gambaryan S, Wiesinger H, Lutzenburg M, Kraemer M, et al. Expression of the Na⁺-D-glucose cotransporter SGLT1 in neurons. *J Neurochem.* 1997;69(1):84–94.
62. Elfeber K, Köhler A, Lutzenburg M, Osswald C, Galla HJ, Witte OW, et al. Localization of the Na⁺-D-glucose cotransporter SGLT1 in the blood-brain barrier. *Histochem Cell Biol.* 2004;121(3):201–7.
63. Vemula S, Roder KE, Yang T, Bhat GJ, Thekkumkara TJ, Abbruscato TJ. A functional role for sodium-dependent glucose transport across the blood-brain barrier during oxygen glucose deprivation. *J Pharmacol Exp Ther.* 2009;328(2):487–95.
64. Newgard CB, An J, Bain JR, Muehlbauer MJ, Stevens RD, Lien LF, et al. A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metabol.* 2009;9(4):311–26.
65. Woo SL, Yang J, Hsu M, Yang A, Zhang L, Lee RP, et al. Effects of branched-chain amino acids on glucose metabolism in obese, prediabetic men and women: a randomized, crossover study. *Am J Clin Nutr.* 2019;109(6):1569–77.
66. Iwao M, Gotoh K, Arakawa M, Endo M, Honda K, Seike M, et al. Supplementa-tion of branched-chain amino acids decreases fat accumulation in the liver through intestinal microbiota-mediated production of acetic acid. *Sci Rep.* 2020;10(1):18768.
67. Furuya F, Fujita Y, Matsuo N, Minamino H, Oguri Y, Isomura N, et al. Liver autophagy-induced valine and leucine in plasma reflect the metabolic effect of sodium glucose co-transporter 2 inhibitor dapagliflozin. *EBioMedicine.* 2022;86:104342.
68. Velloso NA, Dalmolin GD, Fonini G, Gindri Sinhorin VD, Ferreira da Silveira A, Rubin MA, et al. Spermine attenuates behavioral and biochemical alterations induced by quinolinic acid in the striatum of rats. *Brain Res.* 2008;1198:107–14.
69. Shin TH, Phukan G, Shim JS, Nguyen DT, Kim Y, Oh-Lee JD, et al. Restoration of polyamine metabolic patterns in vivo and in vitro Model of ischemic stroke following human mesenchymal stem cell treatment. *Stem Cells Int.* 2016;2016:4612531.
70. Li J, Doyle KM, Tatlisumak T. Polyamines in the brain: distribution, biological interactions, and their potential therapeutic role in brain ischaemia. *Curr Med Chem.* 2007;14(17):1807–13.
71. Bellé NA, Dalmolin GD, Fonini G, Rubin MA, Rocha JB. Polyamines reduces lipid peroxidation induced by different pro-oxidant agents. *Brain Res.* 2004;1008(2):245–51.
72. Xuan M, Gu X, Li J, Huang D, Xue C, He Y. Polyamines: their significance for maintaining health and contributing to diseases. *Cell Communication Signal-ing: CCS.* 2023;21(1):348.
73. Mindikoglu AL, Opekun AR, Putluri N, Devaraj S, Sheikh-Hamad D, Vierling JM, et al. Unique metabolomic signature associated with hepatorenal

- dysfunction and mortality in cirrhosis. *Translational Research: J Lab Clin Med.* 2018;195:25–47.
74. Serena J, Leira R, Castillo J, Pumar JM, Castellanos M, Dávalos A. Neurological deterioration in acute lacunar infarctions: the role of excitatory and inhibitory neurotransmitters. *Stroke.* 2001;32(5):1154–61.
 75. Tong XK, Trigiani LJ, Hamel E. High cholesterol triggers white matter alterations and cognitive deficits in a mouse model of cerebrovascular disease: benefits of simvastatin. *Cell Death Dis.* 2019;10(2):89.
 76. de Oliveira J, Engel DF, de Paula GC, Dos Santos DB, Lopes JB, Farina M, et al. High cholesterol Diet exacerbates blood-brain barrier disruption in LDLr^{-/-} mice: impact on cognitive function. *J Alzheimer's Disease: JAD.* 2020;78(1):97–115.
 77. Engel DF, de Oliveira J, Lopes JB, Santos DB, Moreira ELG, Farina M, et al. Is there an association between hypercholesterolemia and depression? Behavioral evidence from the LDLr^{-/-} mouse experimental model. *Behav Brain Res.* 2016;311:31–8.
 78. Ishikawa M, Stokes KY, Zhang JH, Nanda A, Granger DN. Cerebral microvascular responses to hypercholesterolemia: roles of NADPH oxidase and P-selectin. *Circul Res.* 2004;94(2):239–44.
 79. Lopes FG, Bottino DA, Oliveira FJ, Mecenas AS, Clapauch R, Bouskela E. In elderly women moderate hypercholesterolemia is associated to endothelial and microcirculatory impairments. *Microvasc Res.* 2013;85:99–103.
 80. Rodrigues SF, Almeida-Paula LD, Granger DN. Synergistic effects of high blood cholesterol and hypertension on leukocyte and platelet recruitment in the cerebral microcirculation. *Hypertens (Dallas Tex: 1979).* 2014;63(4):747–52.
 81. Kraft P, Schuhmann MK, Garz C, Jandke S, Urlaub D, Mencl S, et al. Hypercholesterolemia induced cerebral small vessel disease. *PLoS ONE.* 2017;12(8):e0182822.
 82. Liu Y, Yuan C, Chen X, Fang X, Hao J, Zhou M, et al. Association of Plasma Lipids with White Matter hyperintensities in patients with Acute ischemic stroke. *Int J Gen Med.* 2023;16:5405–15.
 83. Yu X, Yu Y, Wei C, Wang L, Jiang J, Zhang R, et al. Association between small dense low-density lipoprotein cholesterol and neuroimaging markers of cerebral small vessel disease in middle-aged and elderly Chinese populations. *BMC Neurol.* 2021;21(1):436.
 84. Kang SH, Yoo H, Cheon BK, Park YH, Kim SJ, Ham H, et al. Distinct effects of cholesterol profile components on amyloid and vascular burdens. *Alzheimers Res Ther.* 2023;15(1):197.
 85. Johnson NF, Gold BT, Ross D, Bailey AL, Clasey JL, Gupta V, et al. Non-fasting high-density lipoprotein is Associated with White Matter Microstructure in healthy older adults. *Front Aging Neurosci.* 2019;11:100.
 86. Zhang Y, Ngo D, Yu B, Shah NA, Chen H, Ramos AR, et al. Development and validation of a metabolite index for obstructive sleep apnea across race/ethnicities. *Sci Rep.* 2022;12(1):21805.
 87. Chokesuwattanaskul A, Lertjitbanjong P, Thongprayoon C, Bathini T, Sharma K, Mao MA, et al. Impact of obstructive sleep apnea on silent cerebral small vessel disease: a systematic review and meta-analysis. *Sleep Med.* 2020;68:80–8.
 88. Burgess S, Davies NM, Thompson SG. Bias due to participant overlap in two-sample mendelian randomization. *Genet Epidemiol.* 2016;40(7):597–608.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.